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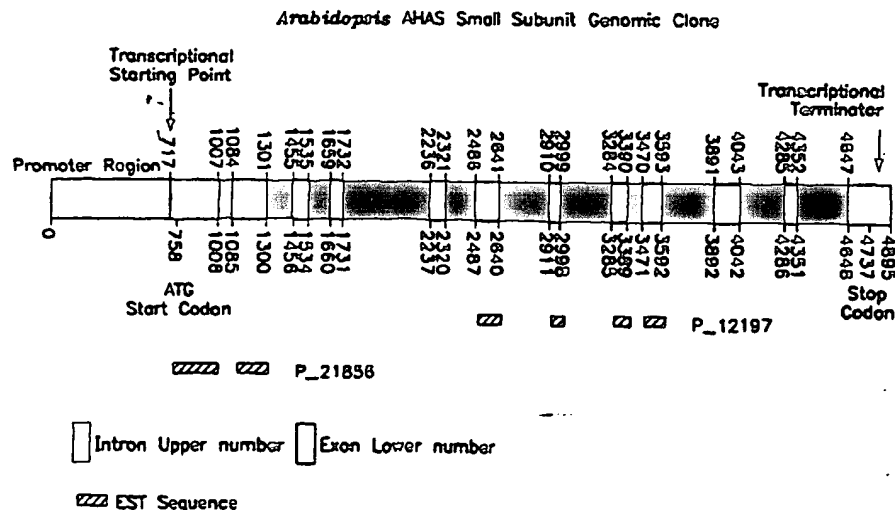
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**(54) Title: GENES AND VECTORS FOR CONFERRING HERBICIDE RESISTANCE IN PLANTS**



**(57) Abstract**

Genomic and cDNA sequences and plant expression vectors encoding for an eukaryotic AHAS small subunit protein are disclosed. The DNA sequences and vectors are used to transform plants to produce transgenic plants which possess elevated levels of tolerance or resistance to herbicides, such as imidazolinones.

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**Description**

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GENES AND VECTORS FOR CONFERRING  
HERBICIDE RESISTANCE IN PLANTS

CROSS REFERENCE TO RELATED APPLICATION

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10 This application claims the benefit, under 35 U.S.C. 119(e), of U.S. Provisional Patent  
Application Serial No. 60/106,239 filed October 29, 1998.

15  
BACKGROUND OF THE INVENTION

10 Herbicides are used extensively in agronomy for controlling weeds and other undesirable  
plants. Because of their phytotoxicity, herbicides also kill or significantly inhibit the growth and  
20 yield of desirable plants.

Some plants, for example *Arabidopsis*, inherently possess or develop resistance to certain  
herbicides upon repeated exposure to herbicides with the same mode of action. It has been a goal  
15 of plant biotechnologists to identify, isolate and clone plant genes that confer resistance to  
25 herbicides and use these genes to transform desirable plants such as crops to render them  
herbicide resistant.

Several methods for generating or identifying herbicide resistance in plants are known.  
30 For example, U.S. Patents Nos. 5,719,046, 5,633,444 and 5,597,717 disclose a plant sulfonamide  
20 resistant gene and methods for transforming plant cells whose growth is inhibited by  
sulfonamides, with vectors containing this gene.

U.S. Patent No. 5,405,765 discloses a method for producing transgenic wheat plants.  
35 This method comprises delivering a heterologous DNA to a Type C embryonic wheat callus in a  
suspension culture by an accelerated particle bombardment method.

25 U.S. Patent No. 5,539,092 discloses polynucleotides encoding a cyanobacterial and plant  
40 acetyl-CoA carboxylase. This patent discloses processes for increasing the herbicide resistance of  
a monocotyledonous plant, comprising transforming the plant with a DNA molecule encoding a  
herbicide resistant polypeptide having the ability to catalyze the carboxylation of acetyl-CoA. The  
patent further discloses that the transgenic plants produced are resistant to herbicides such as  
45 30 arylphenoxypropionates and cyclohexanediones.

U.S. Patent No. 5,304,732 discloses methods for isolating herbicide-resistant plants. The  
patent describes the use of *in vitro* cell culture methods for isolating plant cell lines that are  
50 resistant to herbicides such as imidazolinones and sulfonamides.

5           The trait for a specific herbicide resistance is most often associated with a particular  
enzyme. One such enzyme which has been of interest in its association of conferring herbicide  
resistance in plants is acetohydroxy-acid synthase ("AHAS"), also known as acetolactate synthase  
10 ("ALS," E.C. 4.1.3.18). It is an essential enzyme in plants and many microorganisms, and in most  
5 plants the enzyme is sensitive to herbicides. The AHAS enzyme catalyzes the first step in the  
biosynthesis of the branched-chain amino acids, isoleucine, leucine, and valine, and its activity is  
allosterically inhibited by these amino acids. AHAS activity is also inhibited by several classes of  
15 herbicides, including imidazolinone compounds such as imazethapyr (PURSUIT®, American  
Cyanamid, Parsipanny, NJ); sulfonylurea-based compounds such as sulfometuron methyl  
10 (OUST®, E.I. du Pont de Nemours and Company, Wilmington, DE); triazolopyrimidine  
sulfonamides (Broadstrike™, Dow Elanco; see Gerwick et al. *Pestic. Sci.* 29: 357-364, 1990);  
20 sulfamoylureas (Rodaway et al., *Mechanism of Selectivity of Ac 322,140 in Paddy Rice, Wheat  
and Barley*, Proc. Brighton Crop Protec. Conf., Weeds, 1993); pyrimidyl-oxy-benzoic acids  
(STABLE®, Kumiai Chemical Industry Co., E.I. du Pont de Nemours and Company), and  
25 15 sulfonfylcarboxamides (Alvarado et al., U.S. Patent No. 4,883,914). Inhibition of AHAS activity  
may lead to the inability of the plant to make the branched amino acids or to the accumulation of  
toxic metabolites and, therefore, plant death.

30           Genes encoding AHAS enzymes have been isolated from enteric bacteria, including  
*Escherichia coli*, and *Salmonella typhimurium*. U.S. Patent No. 5,643,779 discloses a nucleic  
20 acid sequence coding for an  $\alpha$ -AHAS enzyme from *Lactococcus* and vectors containing same for  
transforming microorganisms. The transgenic microorganisms produce an enhanced amount of  
35 the AHAS enzyme.

40           Japanese Patent Document No. JP08214882 discloses a nucleic acid sequence for a large  
25 and a small subunit of AHAS from *Rhodobacter capsulatus*. The gene sequences are used to  
transform photosynthetic microorganisms to improve the production of AHAS enzyme for the  
synthesis of amino acids.

45           In eukaryotes, a gene encoding a polypeptide homologous to the large subunits of  
bacterial AHAS enzymes has been identified in the yeast *Saccharomyces cerevisiae*. Genes  
encoding mutant large subunits of AHAS from various plants have also been isolated, cloned, and  
30 used to create transgenic plants that are resistant to herbicides.

50           U.S. Patents Nos. 5,605,011, 5,013,659, 5,141,870 and 5,378,824 disclose nucleic acid  
fragments encoding a mutant plant ALS protein associated with herbicide resistance. The mutant

5 ALS large subunit protein confers herbicide resistance to sulfonylurea compounds in plants. The nucleic acid fragments encoding this mutant large subunit protein are used in vectors to transform plants which are normally sensitive to sulfonylurea herbicides. The transgenic plants resulting from such transformation are resistant to sulfonylurea herbicides.

10 U.S. Patent No. 5,633,437 discloses a large subunit gene and enzyme isolated from cocklebur, *Xanthium* sp., which confer resistance to several structurally unrelated classes of herbicides in plants, plant tissues and seeds. The patent discloses herbicides which normally inhibit AHAS activity.

15 Herbicide resistant AHAS large subunit genes have also been rationally designed. WO 96/33270, U.S. Patent Nos. 5,853,973 and 5,928,937 disclose structure-based modeling methods for the preparation of AHAS variants, including those that exhibit selectively increased resistance to herbicides such as imidazolines and AHAS inhibiting herbicides. This document discloses isolated DNAs encoding such variants, vectors containing these DNAs, methods for producing the variant polypeptides, and herbicide resistant plants containing specific AHAS gene mutations.

20 The prokaryotic AHAS enzymes exist as two distinct, but physically associated, protein subunits. In prokaryotes, the two polypeptides, a "large subunit" and a "small subunit," are expressed from separate genes. Three major AHAS enzymes, designated I, II and III, all having large and small subunits, have been identified in enteric bacteria. In prokaryotes, the AHAS enzyme has been shown to be a regulatory enzyme in the branched amino acid biosynthetic pathway (Mifflin, B.J. *Arch. Biochem. Biophys.* 146:542-550, 1971), and only the large subunit has been observed as having catalytic activity. From studies of AHAS enzymes from microbial systems, two roles have been described for the small subunit: 1) the small subunit is involved in the allosteric feedback inhibition of the catalytic large subunit when in the presence of isoleucine, leucine or valine or combinations thereof; and 2) the small subunit enhances the activity of the large subunit in the absence of isoleucine, leucine or valine. The small subunit has also been shown to increase the stability of the active conformation of the large subunit (Weinstock et al. *J. Bacteriol.* 174:5560-5566, 1992). The expression of the small subunit can also increase the expression of the large subunit as seen for AHAS I from *E. coli* (Weinstock et al. *J. Bacteriol.* 174:5560-5566, 1992).

25 In these microbial systems, the large subunit alone *in vitro* exhibits a basal level of activity that cannot be feedback-inhibited by the amino acids isoleucine, leucine or valine. When the small

5 subunit is added to the same reaction mixture containing the large subunit, the specific activity of the large subunit increases.

10 The large AHAS subunit protein has been identified in plants and isolated and used to transform plants. An AHAS mutant allele isotype of the AHAS3 large subunit protein, having the tryptophan at position 557 replaced with leucine has been found in a *Brassica napus* cell line (Hattori et al. *Mol. & Gen. Genet.* 246: 419-425, 1995). The mutant protein product of this gene confers sulfonylurea, imidazolinone and triazolopyridine resistance to the cell line. This mutant allele, when expressed in transgenic plants, also confers resistance to these herbicides.

15 An AHAS herbicide-resistant, double-mutant allele of the large subunit of *Arabidopsis thaliana* has also been identified (*Planta* 196:64-68, 1995). The gene, *csr1-4*, encodes an AHAS enzyme with altered kinetics which is resistant to chlorsulfuron, imazapyr, and triazolopyrimidine. The *csr1-4* gene when expressed in plants affects the growth of the plants in response to added L-valine and L-leucine.

20 Until recently, there was no direct evidence that a small subunit protein of AHAS existed in eukaryotic organisms. Recently, other groups, through the use of Expressed Sequence Tags (ESTs), have identified sequences homologous to the microbial AHAS small subunit genes in a eukaryote, the plant *Arabidopsis*. They showed that a randomly isolated *Arabidopsis* cDNA sequence had sequence homology with AHAS small subunit sequences from microbial systems. Since then, ESTs from small subunit genes have been described from other eukaryotes such as yeast and red algae (Duggleby 1997, *Gene* 190:245). Duggleby discloses three EST sequences, two from *Arabidopsis* and one from rice, that have homology to known prokaryotic small subunit gene sequences.

25 More recently, WO 98/37206 discloses the use of an ALS small subunit cDNA sequence from *Nicotiana plumbaginifolia* for screening herbicides which inhibit the holoenzyme. Until the present invention, however, the complete genomic sequence of a eukaryotic AHAS small subunit protein gene had not been determined, nor had a eukaryotic AHAS small subunit protein been produced or isolated from *Arabidopsis*.

#### 45 SUMMARY OF THE INVENTION

30 The present invention provides DNA sequences encoding a biologically functional eukaryotic AHAS small subunit protein and functional variants thereof. In accordance with the invention, the *Arabidopsis* AHAS small subunit gene has been cloned and sequenced. Expression vectors containing DNA sequences encoding a eukaryotic AHAS small subunit protein are



5 provided for transforming plants. Expression vectors are also provided which contain genes  
encoding both large and small subunits of an AHAS protein or proteins. The vectors may be used  
in methods to produce transgenic plants of interest, such as dicot and monocot crop-plants,  
10 including wheat, barley, rice, sugarcane, cotton, corn, soybean, sugar beet, canola and the like.  
5 The transgenic plants so produced will possess an elevated level of tolerance to certain herbicides,  
such as imidazolinones.

15 The invention also relates to methods for constructing DNA vectors including plasmids  
containing the AHAS small subunit protein genes. The vectors of the invention are suitable for  
transforming a broad range of plants and may also be engineered to contain a DNA sequence  
10 encoding an AHAS large subunit protein.

20 Vectors containing the complementary eukaryotic small subunit protein gene, for example,  
the gene derived from *Arabidopsis* or maize, can be used to transform an imidazolinone-tolerant  
plant to enhance herbicide resistance by a secondary mechanism.

25 In a specific embodiment of the invention, expression vectors are provided which contain  
15 DNA sequences encoding the large and small AHAS subunit proteins and the promoters for the  
large and small subunits of AHAS as coordinately regulated expression systems in plants.

30 For certain monocot crops, it may be preferred to use a monocot AHAS small subunit  
gene and promoter such as those derived from rice or maize. These genes are useful for  
applications involving the development of transgenic monocot plants which exhibit herbicide  
20 resistance to imidazolinones or other AHAS-inhibiting herbicides.

35 In one embodiment, the invention relates to a method for creating transgenic crop plants  
that exhibit high level tolerance or resistance to imidazolinone herbicides. The method comprises  
introducing a DNA construct, such as a plasmid vector containing an herbicide resistant mutant of  
the AHAS large subunit gene and an AHAS small subunit gene, into a plant that is normally  
40 25 sensitive or partially resistant to imidazolinones. Once the vector is introduced into plant tissue,  
the vector uses the endogenous mechanisms of the plant to express the large and small subunit  
proteins. The increased production of exogenous large and small subunits of the AHAS enzyme  
confers enhanced imidazolinone resistance to the plant. This increased imidazolinone resistance  
45 results from an increase in catalytic activity, stability, resistance to degradation or resistance to  
30 inhibition of the large subunit protein in the presence of increased amounts of small subunit  
protein in the plant.

5 In a preferred embodiment, the large and small subunit genes of the AHAS-enzyme are present on a single plasmid which integrates as one into the genome of the transformed plants, and segregates as a single locus for easier breeding of herbicide resistant crops.

10 In another embodiment of the invention, the DNA construct or vector comprises a herbicide-resistant large AHAS subunit gene and a small AHAS subunit protein gene fused into a single gene, operably linked to and expressed from a single promoter.

15 The small subunit protein of AHAS produced by the present vectors may also be used as a new target site for herbicides or used in combination with the large subunit to screen for putative inhibitors of the large subunit.

20 The invention also relates to methods of utilizing the small subunit DNA sequences as screening tools to identify mutations of the AHAS enzyme which confer herbicide resistance in plants. In this aspect of the invention, organisms coexpressing the large and small subunit of AHAS are screened for mutations which confer resistance to herbicides in plants. The mutant gene products are isolated and tested *in vivo* for the effects of herbicides including  
25 imidazolinones. Then, mutant herbicide resistant forms are isolated.

#### BRIEF DESCRIPTION OF THE DRAWINGS

30 FIG. 1 depicts the pUC19 plasmid plant expression vector construct containing an *Arabidopsis* AHAS small subunit genomic DNA sequences of the invention.

20 FIG. 2 depicts an *Arabidopsis* AHAS small subunit gene map.

35 FIG. 3 depicts a plasmid expression vector, pHUWE82 of the invention, which contains the *Arabidopsis* AHAS small subunit gene without the first three codons.

40 FIG. 4 depicts a plasmid expression vector, pHUWE83 of the invention, which contains the *Arabidopsis* AHAS small subunit gene minus the nucleotide sequence coding for the first 98 amino acids.

45 FIG. 5 is a bar graph showing the *in vitro* activity and stability of the large AHAS subunit protein of the *Arabidopsis* AHAS enzyme in the presence of Phosphate Buffered Saline (MTPBS) and dithiothreitol (DTT).

50 FIG. 6 is a bar graph showing the *in vitro* activity of the large subunit protein of the *Arabidopsis* AHAS enzyme in the presence of Phosphate Buffered Saline (MTPBS).

FIG. 7 is a graph showing the *in vitro* activity of the large subunit protein of the *Arabidopsis* AHAS enzyme in the presence of increasing amounts of bovine serum albumin (BSA).

FIG. 8 is a graph showing the activity *in vitro* of the wild type large subunit protein of the AHAS enzyme in increasing amounts of the *Arabidopsis* small subunit protein.

FIG. 9 is a graph showing the *in vitro* activity of an herbicide resistant mutant (Met124 His) large subunit *Arabidopsis* AHAS protein in the presence of the *Arabidopsis* AHAS small subunit protein.

FIG. 10 is a plant transformation vector, pHUWE67 of the invention which contains a 5.6 kb DNA fragment containing the AHAS small subunit genomic DNA.

FIG. 11A-11E illustrate the plant transformation (expression) vectors of the invention.

#### DETAILED DESCRIPTION OF THE INVENTION

The present invention relates to the cloning and sequencing of an AHAS small subunit protein gene of *Arabidopsis*. SEQ ID NO:1 in the Sequence Listing contains the nucleotide sequence of the *Arabidopsis* AHAS small subunit cDNA. The corresponding amino acid sequence of the encoded AHAS small subunit polypeptide is shown in SEQ ID NO:2 in the Sequence Listing. The genomic DNA sequence of the *Arabidopsis* AHAS small subunit gene is shown in SEQ ID NO:3 of the Sequence Listing.

The invention also relates to an isolated DNA sequence encoding a eukaryotic acetohydroxy-acid synthase, AHAS small subunit protein. Specifically, the invention relates to a DNA sequence which encodes a plant AHAS small subunit protein, which can be obtained from a dicotyledonous plant such as *Arabidopsis*, or a monocotyledonous plant such as rice or maize.

The cloned *Arabidopsis* AHAS small subunit gene sequences can be used in DNA vector constructs to transform crop plants which are normally sensitive or partially resistant to herbicides such as imidazolinone. The transgenic plants obtained following transformation show increased resistance to AHAS-inhibiting herbicides such as imidazolinone.

The expression vector systems of the present invention can be used under suitable conditions to transform virtually any plant cell. Transformed cells can be regenerated into whole plants so that when a gene is expressed in intact plants, it imparts herbicide resistance to the transgenic plant.

5 The DNA sequence encoding the AHAS small subunit may be used in vectors to transform plants so the plants created have enhanced resistant to herbicides, particularly imidazolinones. The DNA sequence encoding the AHAS small subunit protein may be used in  
10 vectors alone or in combination with a DNA sequence encoding the large subunit of the AHAS enzyme in conferring herbicide resistance in plants.

The invention also relates to a plant expression vector comprising a eukaryotic promoter and a DNA sequence encoding a eukaryotic AHAS small subunit protein. The eukaryotic promoter for use in the expression vector should be a high level expression plant promoter, such as the *Arabidopsis* AHAS small subunit promoter. The AHAS small subunit gene sequences  
15 preferably used in the expression vector is a DNA sequence which encodes the *Arabidopsis* small subunit protein.

In another embodiment, the plant expression vector comprises a promoter for the large subunit of eukaryotic AHAS protein; a DNA sequence encoding the large subunit of a eukaryotic AHAS protein; a promoter for the small subunit of the AHAS protein; and a DNA sequence  
20 encoding for a small subunit of a eukaryotic AHAS protein.

In yet another embodiment, the plant expression vector for expressing a heterologous AHAS gene in a plant, comprises a plant promoter and a DNA sequence encoding a fusion protein comprising a large subunit and a small subunit of a eukaryotic AHAS protein.

In one embodiment, the plant expression vector comprises the *Arabidopsis* AHAS small subunit promoter and a DNA sequence encoding an *Arabidopsis* small subunit AHAS protein.  
25

In another embodiment, the plant expression vector comprises in series a promoter expressible in a plant cell; a DNA sequence encoding a transit AHAS large subunit polypeptide; a DNA sequence encoding a wild type, mature AHAS large subunit protein or variant; a DNA sequence encoding a linker polypeptide transcript; a DNA sequence encoding a mature eukaryotic  
30 AHAS small subunit protein; and a plant terminator sequence. In this embodiment, the promoter, the DNA sequence encoding the transit AHAS large subunit protein and the DNA sequence encoding the mature AHAS large subunit protein which are preferably derived from a dicotyledonous plant, particularly from *Arabidopsis*. Alternatively, the plant expression vector may comprise a promoter, a DNA sequence encoding the transit AHAS large subunit protein and  
35 a DNA sequence encoding the mature AHAS large subunit protein are derived from a monocotyledonous plant such as maize.

5 In another embodiment, the plant expression vector comprises a promoter suitable for expression in plants, a DNA sequence encoding a fusion protein comprising a large subunit and a small subunit of a eukaryotic AHAS protein.

10 In another embodiment, the plant expression vector comprises DNA sequences for enhancing gene expression, such as introns and leader sequences. In this aspect of the invention, the plant expression vector comprises a DNA sequence for regulating AHAS gene expression. The intron sequences may also be a heterologous intron sequence from an intron such as the maize *Adhl* intron and the first intron from the shrunkent-1 locus. In addition, the DNA sequences for enhancing gene expression may be a leader sequence such as the W-sequence from the Tobacco Mosaic virus.

20 The invention also relates to an isolated eukaryotic AHAS small subunit protein. The AHAS small subunit protein has the amino acid sequence corresponding to SEQ ID NO: 2 in the Sequence Listing. This protein can be purified from, for example, *Arabidopsis* and may be used in compositions. Also, the gene can be used to express the *Arabidopsis* small subunit protein in a microbe such as *E. coli* and purified from extracts of *E. coli*.

25 The invention also relates to a method for creating a transgenic plant which is resistant to herbicides, comprising transforming a plant with a plant expression vector comprising a DNA sequence encoding a eukaryotic AHAS small subunit protein.

30 The invention also relates to a method for imparting herbicide resistance to a plant cell, comprising co-transforming the plant cell with a first plant expression vector comprising a first plant expressible promoter, and a DNA sequence encoding the large subunit of the AHAS protein and a second plant expression vector comprising a second plant expressible promoter and a DNA sequence encoding the small subunit of an eukaryotic AHAS protein.

35 The invention further relates to a method for enhancing the herbicide resistance of a transgenic plant which expresses a gene encoding an AHAS large subunit protein or a mutant or variant thereof, comprising transforming the transgenic plant with a DNA sequence encoding a eukaryotic small subunit AHAS protein or a mutant or variant thereof.

40 The invention also relates to a method for enhancing the herbicide resistance in the progeny plants of a plant, which comprises somatically or sexually crossing the plant with a transgenic plant whose genetic complement comprises a sequence encoding a herbicide-resistant mutant of the large subunit of a eukaryotic AHAS protein, and a DNA sequence encoding the

5 small subunit of an eukaryotic AHAS protein; and selecting for those progeny plants which exhibit herbicide resistance.

10 The invention also relates to the transgenic plants and progeny produced by the methods of the invention, which plants exhibit elevated resistance to imidazolinone and other herbicides.

15 The invention also relates to a transgenic plant whose genetic complement comprises a plant expressible gene comprising a promoter for expression in plants, a DNA sequence encoding a fusion protein comprising a large subunit and a small subunit of a eukaryotic AHAS protein and a terminator sequence which functions in plant cells.

20 The invention also relates to a method for identifying mutations in the plant AHAS genes which confer resistance to herbicides, comprising exposing an organism to a herbicide compound, which organism possesses a heterologous vector comprising an AHAS small subunit protein gene. In another aspect of this embodiment, the heterologous vector may comprise the AHAS large and the small subunit genes. This method is also useful as a screening system for testing the effects of herbicides on mutant forms of the AHAS enzyme.

25 The invention also relates to a method for identifying mutations in the plant AHAS gene(s) which alters the allosteric feedback inhibition characteristics of the enzyme. Mutations that alter the feedback characteristics of the enzyme, in either the AHAS large or small subunit genes are used to alter amino acid levels in plants, particularly of the branched chain amino acids. The method comprises: transforming a microbial strain which is deficient in AHAS enzyme activity with a plasmid expression vector comprising a mutant plant AHAS small subunit gene. A suitable microbial strain which lacks AHAS activity is *E. coli* M1262. The mutant AHAS large and small subunit genes can be generated randomly or rationally designed from protein structural models using methods previously described (Ott et al. *J. Mol. Biol.* 263: 359-368, 1996). Once the microbial strain is transformed, they are screened in minimal medium in the presence of one or two, but not three branched chain amino acids, and then the microbial strains which grow in the minimal medium are identified.

30 The vectors containing the AHAS small subunit gene can be incorporated into plant or bacterial cells using conventional recombinant DNA technology. Plants are grown from transformed plant cells and second generation plants can be obtained from the seeds of the transgenic plants. Alternatively, the vectors for transforming plants can be recombinant plant viral vectors containing an expressible AHAS small subunit protein gene. In this embodiment, the viral vectors

are capable of systemically infecting the target crop plants and capable of expressing the AHAS small subunit protein in the host plant without disrupting the genome of the host.

The invention also relates to the AHAS small subunit gene promoter DNA sequences. In this aspect of the invention, AHAS small subunit promoter sequences can be used to express heterologous polypeptides. Alternatively, the AHAS small and large subunit promoters can also be used as a coordinately regulated gene system to express heterologous multi-subunit proteins or to overexpress a single gene.

#### Identification, Cloning and Sequencing of the AHAS Small Subunit Protein Gene

The EST sequence of the putative *Arabidopsis thaliana* small subunit protein, designated P\_12197 in the GenBank, was used to clone the complete AHAS small subunit gene. Synthetic polymerase chain reaction (PCR) primers were specifically designed to correspond to the putative AHAS small subunit DNA sequences of *Arabidopsis* corresponding to the AHAS small subunit EST sequences in deposit in the GenBank. The primers were synthesized using standard techniques (U.S. Patent No. 4,683,202; Sambrook et al. Molecular Cloning 2<sup>nd</sup> Ed., Cold Spring Harbor).

Reverse Transcriptase (RT)-PCR was performed on total RNA isolated from *Arabidopsis*. Primers designed from the EST sequence were able to amplify an *Arabidopsis* cDNA fragment. This fragment was cloned into an Invitrogen TA vector (Invitrogen Cat. No. K2000-01) using standard techniques. This clone was named pDGR102 and corresponded to a 450 base-pair fragment containing a portion of the EST sequence.

The same PCR primers were also used to amplify a fragment from an *Arabidopsis*  $\lambda$ :yes cDNA library. This confirmed that an AHAS small subunit gene was present in the library. Using a sense strand primer specific within pDGR102, and a reverse primer that hybridized to the  $\lambda$ :yes phagemid vector, a fragment that represented the 3' half of the small subunit gene was amplified by PCR utilizing the *Arabidopsis* total cDNA library as a template source. This product of approximately 800 bases was cloned into the Invitrogen TA vector (Invitrogen, Cat. No. K2000-01) and named clone pDGR106. Clone pDGR106 was also sequenced, and its translated amino acid sequence confirmed that the fragment represented the 3' half of the small subunit gene by homology to known prokaryotic small subunit gene sequences. This fragment contained the stop codon and 3' flanking poly A tail.

The PCR fragment contained in pDGR102 and pDGR106 were found to represent a 5' region and the 3' half, respectively, of the small subunit gene, and their DNA sequences overlapped

5 by approximately 188 bp. A unique Ssp I restriction enzyme site located in the overlap region  
was used to cleave and ligate the fragment together to reconstruct a nearly full-length AHAS  
small subunit gene (a portion of the N-terminal gene was still missing). The resulting clone was  
10 labeled pDGR115.

5 5' Rapid Amplification of cDNA Ends (5' RACE, GIBCO/BRL Cat. No. 18374-058) was  
used to complete sequencing of the 5' end of the *Arabidopsis* AHAS small subunit gene. Primers  
designed from the pDGR115 sequence were used to clone and extend the sequence to the 5' end  
15 of the small subunit gene. Total RNA extracted from *Arabidopsis* seedling were used as  
template. The sequence was extended 650 base pairs and a putative start codon for the N-  
10 terminal methionine residue was identified. The established full length sequence was used to  
generate a full length cDNA clone.

A genomic clone to the *Arabidopsis* small subunit gene was obtained by screening a  
Clonotech *Arabidopsis* genomic lambda library. To screen the library, a 380 bp probe was  
25 generated by PCR amplification of a 5' region of the small subunit cDNA gene sequences. The  
15 PCR product was obtained by using the primers; 5'-CAGAGATCATGTGGCTAGTTGA-3' (SEQ  
ID NO: 4 in the Sequence Listing) and 5'-GAGCGTCGAGAATACGATGTAC-3' (SEQ ID NO:  
5 in the Sequence Listing). The 380 base pair PCR product was cloned into the Invitrogen TA  
cloning vector. To label the probe the PCR insert was cut out by Eco RI and labeled with  $\alpha$ -<sup>32</sup>P  
30 dCTP by random priming. Screening of the library was performed on nylon membranes by  
20 conventional methods. A lambda phage hybridizing to the probe was identified and isolated. The  
lambda phage DNA was extracted and digested with Sal I and the fragments were cloned into  
35 pUC19. A primer specific to the small subunit cDNA sequence was used in sequencing reactions  
with the various cloned Sal I genomic fragment in order to identify the clone containing the small  
subunit gene. A clone containing AHAS small subunit sequence within a 5.6 kb Sal I fragment  
40 25 was identified and it is illustrated within the pMSg6 plasmid in FIG. 1. The promoter region, the  
transit sequence, the mature coding sequence of the small subunit gene, the introns, and the  
translational terminator were identified by sequencing 4.9 kb of the genomic fragment.

45 Through comparison of the genomic and cDNA sequences the start codon for the N-  
terminal methionine was identified. The cDNA for the small subunit gene codes for a polypeptide  
30 of 491 amino acids.

FIG. 2 is a map of the genomic DNA sequences of the *Arabidopsis* AHAS small subunit  
50 gene. As shown in FIG. 2, and referring to SEQ ID NO: 3 in the Sequence Listing, this gene



5 contains a promoter which extends from nucleotide number 1 to nucleotide 757. The start codon  
for the gene corresponds to nucleotides 758-760. The gene contains 11 introns and 12 exons.  
Exon 1 extends from nucleotide 758 to nucleotide 1006. Intron 1 extends from nucleotide 1007  
10 to nucleotide 1084. Exon 2 extends from nucleotide 1085 to nucleotide 1300. Intron 2 extends  
5 from nucleotide 1301 to nucleotide 1455. Exon 3 extends from nucleotide 1456 to nucleotide  
1534. Intron 3 extends from nucleotide 1535 to nucleotide 1659. Exon 4 extends from  
1660 to nucleotide 1731. Intron 4 extends from nucleotide 1732 to nucleotide 2236.  
15 Exon 5 extends from nucleotide 2237 to nucleotide 2320. Intron 5 extends from nucleotide 2321  
to nucleotide 2486. Exon 6 extends from nucleotide 2487 to nucleotide 2640. Intron 6 extends  
10 from nucleotide 2641 to nucleotide 2910. Exon 7 extends from nucleotide 2911 to nucleotide  
2998. Intron 7 extends from nucleotide 2999 to nucleotide 3284. Exon 8 extends from  
20 nucleotide 3285 to nucleotide 3389. Intron 8 extends from nucleotide 3390 to nucleotide 3470.  
Exon 9 extends from nucleotide 3471 to nucleotide 3592. Intron 9 extends from nucleotide 3593  
to nucleotide 3891. Exon 10 extends from nucleotide 3892 to nucleotide 4042. Intron 10  
25 15 extends from nucleotide 4043 to nucleotide 4285. Exon 11 extends from nucleotide 4286 to  
nucleotide 4351. Intron 11 extends from nucleotide 4352 to nucleotide 4647. Exon 12 extends  
from nucleotide 4648 to the stop codon at nucleotide 4737. The transcriptional terminator is  
30 located in the DNA segment between the stop codon at nucleotide 4737 and nucleotide 4895.

The amino acid sequence of the *Arabidopsis* AHAS small subunit protein encoded by the  
20 DNA sequence as described above, has high homology to amino acid sequences of AHAS small  
subunit proteins from prokaryotic organisms. Homology is particularly high in conserved regions  
35 of AHAS small subunit sequences in prokaryotes. As an example, the *Arabidopsis* AHAS small  
subunit gene sequence of the present invention had 42.5% sequence identity to that of the  
*Bacillus subtilis* small subunit gene. This indicated that the genomic clone DNA sequences was  
40 25 that of the *Arabidopsis* AHAS small subunit.

FIG. 2 also shows the location of two Genbank EST sequences with accession numbers  
P\_12197 and P\_21856. Both EST sequences had previously been identified to have homology to  
45 microbial AHAS small subunit genes, suggesting there were two isozymes in *Arabidopsis*. The  
EST sequence from P\_12197 was used to clone the AHAS small subunit cDNA and genomic  
30 clones of the invention. Analysis of the completed AHAS sequences indicated the gene codes for  
two repetitive amino acid sequences with homology to known AHAS small subunits. The AHAS  
50 gene sequences are ordered in tandem within the single polypeptide. After comparing the AHAS

5 small subunit gene sequences with the original two ESTs, it was determined that the two ESTs are part of the same gene, each corresponding to similar regions within each repeated sequence.

Some specific Materials and Methods used for cloning the genomic small subunit gene.  
10 Arabidopsis genomic library - Arabidopsis genomic library was bought from Clontech.

#### 5 Construction of Plasmids Containing the AHAS Small Subunit Gene

DNA molecules containing the AHAS small subunit gene comprising the nucleotide  
15 sequence in SEQ ID NO:1 or SEQ ID NO:3 in the Sequence Listing, or a functional variant thereof, can be inserted into a suitable heterologous expression vector system in proper orientation and correct reading frame. Numerous vector systems, such as plasmids, bacteriophage  
10 viruses and other modified viruses, can be used in practicing the invention. Suitable plasmid vectors include, but are not limited to, pBR322, pUC8, pUC9, pUC18, pUC19, pBI122, pKC37,  
20 pKC101 and TA cloning vectors. Viral vectors such as  $\lambda$ gt10,  $\lambda$ gt11 and Charon 4 can also be used.

#### 15 Construction of F1, F2, F3, pHUWE82, and pHUWE83 Plasmid Expression Vectors

In the present invention, AHAS small subunit cDNA sequences were inserted into a  
30 pGEX-2T or pGEX-4T-2 *E. coli* expression vector obtained from Pharmacia. Five different DNA fragments containing the AHAS small subunit gene sequences were cloned into the pGEX-2T or pGEX-4T-2 expression vectors. These clones were designated F1, F2, F3, pHUWE82, and  
20 pHUWE83 all differing in the amount of coding sequence contained within the expression vector. Plasmids F1, F2 and F3 contain the AHAS small subunit cDNA in pGEX-4T-2 *E. coli* expression  
35 vector and are described in more detail in Example 2 below. The AHAS small subunit cDNA in plasmids pHUWE82 and pHUWE83 was cloned in pGEX-2T *E. coli* expression vectors. The  
40 pHUWE82 vector contained a near full length *Arabidopsis* AHAS small subunit gene (without the first 3 amino acids). pHUWE83 was engineered to express the small subunit gene without the  
25 putative transit sequence (without the first 98 amino acids). A map of the plasmids pHUWE82 and pHUWE83 are shown in FIGs. 3 and 4, respectively. The versions of the small subunit gene  
45 are expressed in *E. coli* as a glutathione transferase/AHAS small subunit fusion protein. After affinity purification of the fusion protein the respective proteins are cleaved by thrombin. Due to  
30 the incorporation of the five amino acid thrombin cleavage site, i.e., Leu-Val-Pro-Arg-Gly-Ser-  
50 (SEQ ID NO:6 in the Sequence Listing), and the location of protease cleavage, an additional

glycine and serine residue is maintained on the N-terminal of the small subunit protein. The resulting AHAS small subunit protein from pHUWE82 has the N-terminal sequence Gly-Ser-Ile-Ser-Val-Ser (SEQ ID NO:7 in the Sequence Listing; the first 3 amino acids Met-Ala-Ala were not incorporated into the vector), and the protein from pHUWE83 has the N-terminal sequence Gly-Ser-Met-Ile-Asn-Arg (SEQ ID NO:8 in the Sequence Listing; the first 98 amino acids were not incorporated into the vector). In both pHUWE82 and pHUWE83 the N-terminal sequence amino acids Gly-Ser- are remnants of the thrombin cleavage site.

#### Construction of Plant Transformation/Expression Vectors

Numerous plant transformation vectors and methods for transforming plants are available

- (An, G. et al. *Plant Physiol.*, 81:301-305, 1986; Fry, J., et al. *Plant Cell Rep.* 6:321-325, 1987; Block, M. *Theor. Appl. Genet.* 76:767-774, 1988; Hinchey, et al. *Stadler. Genet. Symp.* 203:212-212, 1990; Cousins, et al. *Aust. J. Plant Physiol.* 18:481-494, 1991; Chee, P.P. and Slightom, J.L. *Gene* 118:255-260, 1992; Christou, et al. *Trends. Biotechnol.* 10:239-246, 1992; D'Halluin, et al., *Bio/Technol.* 10:309-314, 1992; Dhir, et al. *Plant Physiol.* 99:81-88, 1992; Casas et al. *Proc. Nat. Acad. Sci. USA* 90:11212-11216, 1993; Christou, P. *In Vitro Cell. Dev. Biol. - Plant*; 29P:119-124, 1993; Davies, et al. *Plant Cell Rep.* 12:180-183, 1993; Dong, J.Z. and Mchughen, A. *Plant Sci.* 91:139-148, 1993; Franklin, C.I. and Trieu, T.N. *Plant. Physiol.* 102:167, 1993; Golovkin, et al. *Plant Sci.* 90:41-52, 1993; Guo Chin Sci. Bull. 38:2072-2078, 1993; Asano, et al. *Plant Cell Rep.* 13, 1994; Ayeres N.M. and Park, W.D. *Crit. Rev. Plant. Sci.* 13:219-239, 1994; Barcelo, et al. *Plant. J.* 5:583-592, 1994; Becker, et al. *Plant. J.* 5:299-307, 1994; Borkowska et al. *Acta. Physiol. Plant.* 16:225-230, 1994; Christou, P. *Agro. Food. Ind. Hi Tech.* 5: 17-27, 1994; Eapen et al. *Plant Cell Rep.* 13:582-586, 1994; Hartman, et al. *Biol. Technology* 12:919923, 1994; Ritala, et al. *Plant. Mol. Biol.* 24:317-325, 1994; Wan, Y.C. and Lemaux, P.G. *Plant Physiol.* 104:3748, 1994; Weeks, et al. *J. Cell Biochem*:104, 1994). The AHAS small subunit DNA sequences are inserted into any of the vectors using standard techniques. The selection of the vector depends on the preferred transformation technique and the target plant species to be transformed. In a preferred embodiment, the AHAS small subunit gene can be cloned behind a high level expressing plant promoter, and this construct can then be introduced into a plant that is already transgenic or a plant to be transformed with an herbicide resistant mutant allele of the large subunit protein. In this manner, the effectiveness of the herbicide resistance gene may be enhanced by stabilization or activation of the large subunit protein. This

5 method may be applied to any plant species; however, it is most beneficial when applied to important crops.

Methodologies for constructing plant expression cassettes and introducing foreign DNA  
10 into plants is generally described in the art. For example, foreign DNA can be introduced into  
5 plants, using tumor-inducing (Ti) plasmid vectors. Other methods utilized for foreign DNA delivery involve the use of PEG mediated protoplast transformation, electroporation, microinjection  
whiskers, and biolistics or microprojectile bombardment for direct DNA uptake. Such methods  
15 are known in the art. (U.S. Patent No. 5,405,765 to Vasil et al.; Bilang et al. (1991) *Gene* 100:  
247-250; Scheid et al., (1991) *Mol. Gen. Genet.* 228: 104-112; Guerche et al., (1987) *Plant*  
10 *Science* 52: 111-116; Neuhauser et al., (1987) *Theor. Appl. Genet.* 75: 30-36; Klein et al., (1987)  
*Nature* 327: 70-73; Howell et al., (1980) *Science* 208:1265; Horsch et al., (1985) *Science* 227:  
20 1229-1231; DeBlock et al., (1989) *Plant Physiology* 91: 694-701; *Methods for Plant Molecular*  
*Biology* (Weissbach and Weissbach, eds.) Academic Press, Inc. (1988) and *Methods in Plant*  
*Molecular Biology* (Schuler and Zielinski, eds.) Academic Press, Inc. (1989). The method of  
25 15 transformation depends upon the plant cell to be transformed, stability of vectors used, expression  
level of gene products and other parameters.

The components of the expression cassette may be modified to increase expression of the  
30 inserted gene. For example, truncated sequences, nucleotide substitutions or other modifications  
may be employed. DNA sequences for enhancing gene expression may also be used in the plant  
20 expression vectors. These include the introns of the maize *Adh1*, intron1 gene (Callis et al. *Genes*  
and *Development* 1:1183-1200, 1987), and leader sequences, (W-sequence) from the Tobacco  
35 Mosaic virus (TMV), Maize Chlorotic Mottle Virus and Alfalfa Mosaic Virus (Gallie et al.  
*Nucleic Acid Res.* 15:8693-8711, 1987 and Skuzeski et al. *Plant Molec. Biol.* 15:65-79, 1990).  
The first intron from the shrunken-1 locus of maize, has been shown to increase expression of  
40 25 genes in chimeric gene constructs. U.S. Patent Nos. 5,424,412 and 5,593,874 disclose the use of  
specific introns in gene expression constructs, and Gallie et al. (*Plant Physiol.* 106:929-939,  
1994) also have shown that introns are useful for regulating gene expression on a tissue specific  
45 basis. To further enhance or to optimize AHAS small subunit gene expression, the plant  
expression vectors of the invention may also contain DNA sequences containing matrix  
30 attachment regions (MARs). Plant cells transformed with such modified expression systems,  
then, may exhibit overexpression or constitutive expression of the AHAS small subunit gene.

5 To obtain efficient expression of the AHAS small subunit gene and other genes of the  
present invention, a promoter must be present in the expression vector. Depending upon the host  
cell system utilized, any one of a number of suitable promoters can be used. Suitable promoters  
10 should be high level expression plant promoters which include ubiquitin, nos promoter, the small  
5 subunit ribulose biphosphate carboxylase gene promoter, the small subunit chlorophyll A/B bind-  
ing polypeptide promoter, the 35S promoter of cauliflower mosaic virus, the AHAS large and  
small subunit promoters, (OCS)3 MAS and promoters isolated from plants and plant viruses. See  
15 C.E. Vallejos, et al., "Localization in the Tomato Genome of DNA Restriction Fragments  
Containing Sequences Homologous to the rRNA (45S), the major chlorophyll <sub>A/B</sub> binding  
10 Polypeptide and the Ribulose Bisphosphate Carboxylase Genes," *Genetics* 112: 93-105 (1986).  
Another promoter suitable for transforming plants is the actin promoter. A preferred promoter of  
the invention is the AHAS small subunit promoter for use with the AHAS small subunit gene  
sequences. The expression vector can then be used to transform a plant cell. Other plant tissues  
suitable for transformation include leaf tissues, root tissues, meristems, cultured plant cells such as  
25 calluses, and protoplasts.

Use of the AHAS small subunit gene promoter: In a preferred embodiment, the promoter  
to be used to transcribe the AHAS small subunit gene should be a strong plant promoter. The  
30 native promoter of the AHAS small subunit gene in plants may be used for expressing the AHAS  
small subunit protein gene in transgenic plants. The small subunit gene promoter can also be used  
20 in vectors to express heterologous genes. The AHAS small subunit promoter can also be used in  
vectors in conjunction with the promoter of the AHAS large subunit gene. These promoters,  
35 which would drive the transcription of the two genes coding for two subunits of a single  
multimeric protein, may be utilized as coordinately regulated promoters. For example, both  
promoters may be upregulated simultaneously at a specific time or in a specific tissue such as  
40 25 meristems. The advantage of two different, but simultaneously, active promoters is that they may  
not be susceptible to co-suppression. Co-suppression can occur when two genes of similar  
sequence are present within a transformed organism. Co-suppression causes the same level of  
silencing of expression of the genes.

45 Moreover, a transformation vector containing two genes regulated with the same  
30 promoter sequence can undergo recombination between like sequences, thereby inactivating one  
or both genes. Use of different promoters that are co-regulated may allow for expression of two  
50 genes without problems of recombination, and facilitates the expression of multimeric proteins.

5 The promoter of the AHAS small subunit gene can be used for additional purposes. First, the large and the small subunit genes code for polypeptides that work in concert and in physical contact with each other, the two genes may be coordinately regulated in expression. Having both the large and small subunit promoter may enable the expression of other multimeric proteins, or  
10 overexpression of the same gene from two different promoters. This is advantageous since expressing two genes with the same promoter may cause problems due to recombination of homologous promoter sequences. If two different promoters are used, genes for multimeric proteins may not be expressed at the same time or in the same tissue. Coordinately regulated but heterologous promoters would overcome these problems.

15 Secondly, having both the large and small subunit promoter provides tools to understand gene regulation. These promoters can be ligated to reporter genes to test for determining the coordination of the level, tissue specificity, and coordination of subunit genes.

20 Thirdly, it is advantageous to express the small subunit gene on its own promoter so that it is expressed at the appropriate time and tissue to have the most effect in enhancing herbicide  
25 resistance.

30 Lastly, having two promoters that may be coordinately regulated provides us with a tool for analyzing, and isolating regulatory factors that may be common to each of the promoters. Such factors include transcription factors that regulate expression from both AHAS large and small subunit promoters. The promoter sequences may also have common motifs that may be  
35 involved in coordinate regulation of the two genes. Moreover, the role of introns in the small subunit genomic clone may be involved in co-regulation of the two promoters. The two promoters and the introns provide us with tools to elucidate the mechanism of coordinate regulation of promoters.

#### Use of introns of AHAS small subunit gene

40 25 The genomic clone has several introns which may be used to regulate gene expression. Introns have been shown to regulate gene expression. For example the maize *Adh1* intron 1 significantly increases expression of reporter genes in maize (Callis et al. 1987, Genes & Development 1:1183-1200 by Cold Spring Harbor Laboratory ISSN 0890-9369/87). The first  
45 intron of the shrunken-1 locus of maize has also been shown to increase expression in chimeric gene constructs. U.S. Patent Nos. 5,424,412 and 5,593,874 disclose the use of specific introns  
50 for regulating gene expression.

Introns have also been shown to regulate the level of expression on a tissues specific basis. Gallie et al. (Plant Physiol. 106:929-939) showed that enhancement of gene expression by introns is dependent on cell type. Therefore, the AHAS small subunit gene introns can be used to express genes with particular emphasis in specific tissue types.

It is also advantageous to express the small subunit gene with all of its introns so that it is expressed at the appropriate time and tissue to have the most effect in enhancing herbicide resistance.

#### Use of the AHAS small subunit gene in expression vectors

*Tethered enzyme.* In this embodiment of the invention, the AHAS small subunit is translationally coupled to the large subunit via a transcript coding for a linker polypeptide, such as polyglycine (polyGly). The length of the linker polypeptide tether is varied. The positioning of the two AHAS subunits with respect to the linker polypeptide tether is the large subunit transit sequence followed by the large subunit mature coding sequence, the linker polypeptide transcript, and the small subunit mature coding sequence. An alternative positioning involves switching the mature coding sequences of the large and small subunits about the linker polypeptide transcript with the small subunit transit sequence.

*Tethered enzymes to enhance activity and herbicide resistance.* It has been shown with the *E. coli* enzyme that the association between large and small subunits is loose. It was estimated that in *E. coli* at a concentration of  $10^{-7}$  M for each subunit, the large subunits are only half associated as the  $\alpha_2\beta_2$  active holoenzyme (Sella et al. 1993, *J. Bacteriology* 175:5339-5343). Greatest activity is achieved in molar excesses of the AHAS small subunit protein. Since it has been determined that the AHAS enzyme is most stable and active when both subunits are associated (Weinstock et al 1992, *J. Bacteriology*, 174:5560-5566, Sella et al. 1993, *J. Bacteriology* 175:5339-5343) a highly active and stable enzyme may be created by fusing the two subunits into a single polypeptide. Tethered polypeptides have been shown to function properly. Gilbert et al. expressed two tethered oligosaccharide synthetic enzymes in *E. coli* to produce an enzyme that was functional, stable *in vitro*, and soluble (Gilbert et al. *Nature Biotechnology* 16: 769-772, 1998).

Expression of both the large and small subunits of AHAS as a single polypeptide from a single gene also has advantages for producing transgenic herbicide resistant crops. The use of a single gene to transform and breed plants into elite crop lines is easier and more advantageous than when two or more genes are used.

5           *Fused enzyme pair.* In this aspect of the invention, the AHAS small subunit is positioned in the plant vector directly downstream of the large subunit under the direction of a single promoter. Alternatively, the small and large subunit genes of AHAS can be separated and put under the direction of different promoters within a single construct.

10           5           *Two genes, one construct.* In another aspect of the invention, in this expression vector, both the large and the small subunit of AHAS are placed under the control of separate promoters, in a single plasmid construct. This enables the expression of both genes as separate entities; however, the tandem would behave in the plant progeny as a single locus.

15                   *Two genes, one promoter.* The maize streak virus promoter is a bi-functional promoter able to express genes in two direction. Using this promoter, gene transcription can be initiated on genes coded on opposite strands in the vector and in opposite directions. Therefore, a large and a small subunit genes of AHAS can be expressed from a single promoter.

20                   *Two genes, two constructs.* In another approach, two separate vector constructs are made, each containing either the large or small subunit under the direction of different promoters.  
25           15           This approach requires that the plant be doubly transformed.

                  The plant expression vector should also contain a suitable transcription terminator sequence downstream from the gene sequences. A variety of transcriptional terminators are known for use in plant expression cassettes for correct termination of gene transcription and polyadenylation of the transcript. Terminators for use in the invention include CaMV 35S  
30                   terminator, the nopaline synthase terminator, the pea *bcs* terminator, the *tm1* terminator, the AHAS large and small subunit terminators. These terminators can be used in vectors for use in both monocotyledon and dicotyledon transformation.

                  The gene products of the invention may also be targeted to the chloroplasts. This is accomplished by introducing a signal sequence which can be fused into the gene and thus into the expression vector. The signal sequence which will correspond to the amino terminal end of the gene product (see Comai et al. *J. Biol. Chem.* 263:15104-15109, 1988) is fused into the upstream  
40           25           5' end of the gene. The AHAS small subunit protein of the invention has been found to possess a signal sequence, and therefore, this signal sequence can be used in the vectors of the invention.  
45                   Other signal sequences for use in the invention are known, such as those from the 5' end of cDNAs from the AHAS large or small subunit, CAB protein, the EPSP synthase, the GS2 protein, and the like (Cheng & Jogendorf, *J. Biol. Chem.* 268:2363-2367, 1993).  
50



5 Bacteria from the genus *Agrobacterium* can be utilized to introduce foreign DNA and transform plant cells. Suitable species of such bacterium include *Agrobacterium tumefaciens* and *Agrobacterium rhizogenes*. *Agrobacterium tumefaciens* (e.g., strains LBA4404 or EHA105) is particularly useful due to its well-known ability to transform plants.

10 5 Another approach to transforming plant cells with a heterologous gene involves propelling inert or biologically active particles at plant tissues and cells. U.S. Patent Nos. 4,945,050; 5,036,006; and 5,100,792 all to Sanford et al. disclose this technique. In summary, this procedure involves propelling inert or biologically active particles at the cells under conditions effective to penetrate the outer surface of the cell in such manner as to incorporate the vectors into the interior of the cells. When inert particles are utilized, the vector can be introduced into the cell by coating the particles with the vector containing the desired gene. Alternatively, the target cell can be surrounded by the vector so that the vector is carried into the cell by the wake of the particle. Other biologically active particles including dried yeast cells, dried bacteria, or bacteriophages, each containing the desired DNA, can also be propelled into plant cell tissue. In addition, the vectors of the invention can be constructed so that they are suitable for use in plastid transformation methods using standard techniques.

15 20 The isolated AHAS small subunit gene of the present invention can be utilized to confer herbicide resistance to a wide variety of plant cells including monocots and dicots. Although the gene can be inserted into any plant cell falling within these broad classes, it is particularly useful in crop plant cells, such as those of rice, wheat, barley, rye, corn, carrot, sugarcane, tobacco, bean, pea, soybean, sugar beet, and canola.

25 30 The expression system of the present invention can be used to transform virtually any crop plant cell under suitable conditions. Transformed cells can be regenerated into whole plants such that the AHAS small subunit gene imparts or enhances herbicide resistance to the intact transgenic plants. As set forth above, the expression system can be modified so that the herbicide resistance gene is continuously or constitutively expressed.

35 40 Use of the AHAS small subunit gene to enhance herbicide resistance in plants: A plasmid that contains both the genes for the AHAS large and small subunit can be constructed. In this manner, the two genes would segregate as a single locus making breeding of herbicide resistant crops easier. Alternatively, the large and small subunit can be fused into a single gene expressed from a single promoter. The fusion protein would have elevated levels of activity and herbicide resistance. The large subunit of AHAS may be of a wild type sequence (if resistance is conferred

5 in the presence of an independent or fused small subunit), or may be a mutant large subunit that in  
itself has some level of resistance to herbicides. The presence of the small subunit will 1) enhance  
the activity of the large subunit, 2) enhance the herbicide resistance of the large subunit, 3)  
10 increase the stability of the enzyme when expressed *in vivo* and 4) increase resistance to large sub-  
unit to degradation. The small subunit would in this manner elevate the resistance of the plant/  
crop to the imidazolinone or other herbicides. The elevated resistance would permit the applica-  
tion and/or increase the safety of weed-controlling rates of herbicide without phytotoxicity to the  
15 transformed plant/crop. Ideally, the resistance conferred would elevate resistance to  
imidazolinone and other classes of herbicides without increasing, or by increasing to a lesser  
20 degree, resistance to other AHAS inhibiting herbicides such as sulfonylureas, triazolopyrimidines,  
etc.

Additional aspects about enhancing herbicide resistance by the addition of the small subunit gene  
to plants expressing a herbicide resistant form of the large subunit gene.

25 It has been shown in many cases that mutations in proteins can cause instability, decreases  
15 in activity, and a greater propensity to degradation. Herbicide resistant AHAS genes, particularly  
those from plants, generally contain a mutation that confers the resistance to inhibition by the  
herbicide. This places a greater level of importance in stabilizing, maintaining activity, and resis-  
tance to degradation of these proteins. The accompaniment of the small subunit gene to the large  
30 subunit gene may assist in these areas of susceptibility.

20 Subunits of multi-subunit proteins that are present in the absence or non-stoichiometric  
35 levels of the complementary subunits can be preferentially degraded. This has been shown for the  
large and small subunits of ribulosebiphosphate carboxylase/oxygenase (Spreitzer et al. *Proc.*  
*Natl. Acad. Sci. USA* 82: 5460-5464). If the large subunit gene for AHAS is transformed and  
expressed in a crop that does not have the complementary small subunit, or is expressed at high  
40 25 levels beyond the expression levels of the small subunit gene, the large subunit protein could be  
unstable and preferentially degraded. This could result in a lower level of herbicide resistance.

The association of large and small subunits appears to be highly specific. *E. coli* has three  
45 isozymes of large subunits and three isozymes of small subunits. Each large subunit isozyme  
specifically associates with only one of the small subunit isozymes, even though all subunits are  
30 expressed in the same organism (Weinstock et al 1992, *J. Bacteriology*, 174:5560-5566). This  
specificity suggests that endogenous AHAS small subunit proteins could not stabilize or enhance  
50 the activity of an introduced AHAS large subunit if the large subunit gene is derived from a differ-

ent organism or isozyme pair from that of the small subunit. This places an importance, for purposes of herbicide resistance, in introducing both the large and small subunit genes from the same organism and isozyme pair. The expression of a small subunit gene may ameliorate these problems.

The AHAS small subunit gene in combination with the AHAS large subunit gene can also be used as a marker for selecting transformant plant cells and tissues. Any gene of interest can be incorporated in vectors containing the AHAS large and small subunit genes. The vectors can be introduced into plant cells or tissues that are susceptible to AHAS-inhibiting herbicides. The transformants containing these vectors can be selected in the presence of herbicides using standard techniques.

#### Example 1

DNA and Lambda DNA isolation: DNA isolation was carried out using QIAGEN Spin Miniprep Kit (50) (QIAGEN Cat. No. 27104) and standard procedures as provided by the manufacturer. For Lambda DNA isolation, the QIAGEN Lambda Midi Kit (25) (QIAGEN Cat. No. 12543) was used following the manufacturer's protocol. In TA Cloning, the Invitrogen Original TA Cloning Kit (Invitrogen Cat. No. K2000-01) was used and standard protocols were followed.

Subcloning: Lambda DNA was digested with the appropriate restriction enzyme and mixed with pUC19 which was digested with the same restriction enzyme. After phenol extraction, the insert was ligated with pUC19 by adding 1 µl DNA ligase (4 units/mL) and incubate at 17°C overnight.

5' RACE: The 5' RACE used in the experiments was 5' RACE System for Rapid Amplification of cDNA Ends and was obtained from GIBCO/BRL (Cat. No. 18374-058). The reactions were carried out following standard procedures provided by the manufacturer.

Screening library: The Clontech *Arabidopsis* lambda genomic library was plated at a density of 30,000 plaques/150mm plate as described in the protocol supplied by the manufacturer.

Amersham Nucleic Acid Transfer Membranes Hybond™-N+ (DISC: 0.137m DIA, Removal Rating: 0.45um) were used. The nylon transfer membranes were carefully placed onto the plate surface, and membranes and agar were marked using a sterile needle. The first membrane was removed after 3 minutes and a duplicate membrane was placed on the plate surface and removed after 8 minutes. The membranes were placed, colony side up, on a pad of absorbent filter paper soaked in denaturing buffer (0.5N NaOH, 1.5N NaCl) for 5 minutes, then each membrane was

placed, colony side up, on a pad of absorbent filter paper soaked in neutralizing solution (1M Tris-HCl, 1.5N NaCl) for 5 minutes. The membranes were washed briefly in 2X SSC, and transferred to dry filter paper. The sample was fixed to the membranes by UV crosslinking and vacuum-baked at 80°C for 1 hour. Then the membranes were prehybridized in a buffer containing 50% formamide, 2X SSC, 5X Denhardt's solution, 1% sodium dodecyl sulfate (SDS), 0.05 mg/ml denatured salmon sperm DNA, and 0.05 % NaPPi at 42°C for 2 hour. DNA was digested with restriction enzyme and fractionated on a 1% agarose gel. The DNA fragment containing the AHAS small subunit gene was purified using QIAquick Gel Extraction Kit (50) (QIAGEN Cat. No. 28704). The GIBCO/BRL Life Technologies Random Primers DNA labeling System (Cat. No. 18187-013) was used to label the DNA with the following modification. 125ng of probe DNA was dissolved in 55 µL of distilled water in a microcentrifuge tube and denatured by heating for 5 minutes in a boiling water bath, and immediately cooled on ice. Then, dATP, dGTP, dTTP, [ $\alpha$ - $^{32}$ P]dCTP and Klenow enzyme were added to the denatured DNA, and the mixture was incubated at 25°C for an hour. One volume of formamide was added to the mixture and the reaction was heated at 65°C for 30 minutes. The reaction containing the labeled DNA was added to prehybridization solution and the membranes were hybridized at 42°C for 20 hours with slow shaking. After hybridization, the membranes were washed twice with 0.4X SSC buffer containing 0.1% SDS at room temperature for 10 minutes, followed by a single wash in 0.2X SSC buffer containing 0.1% SDS at 65°C for 30 minutes. The membranes were exposed to X-ray film overnight. Plaques containing DNA which hybridized to the DNA probe on duplicate membranes produced a positive result, and these plaques were isolated. The procedure was duplicated until single isolates could be collected.

Sequencing: The sequencing reaction was carried out using the ABI PRISM DNA sequencing Kit following the ABI protocol provided. After ethanol precipitation, the DNA was dissolved in ABI PRISM Template Suppression Reagent and denatured at 90°C for 5 minutes. Then, the samples were loaded onto an ABI 310 sequencer.

## EXAMPLE 2

### *Preparation of Plasmid DNA containing AHAS Large Subunit Protein Genes*

The wild type (pAC774) and Met92His mutant (pAC786) AHAS large subunit genes from *Arabidopsis* were constructed into the *E. coli* expression vector pGEX-4T-2 from Pharmacia.

5 The genes were constructed to express a glutathione transferase/AHAS large subunit fusion protein to aid in purification, similarly as described for plasmids pHUWE82 and pHUWE83 as described above. A five amino acid protease cleavage site was encoded at the junction of the two  
10 proteins so that they could be cleaved apart after purification. Three vector constructs containing the cDNA sequences of the *Arabidopsis* AHAS small subunit gene were made using the pGEX-4T-2 expression vector. These were designated F1, F2 and F3, all three differing in the amount of peptide sequence contained within the gene. The N-terminal amino acid sequence of the peptide encoded by the AHAS cDNA of the F1 plasmid is Gly-Ser-Pro-Lys-Ile-Ala-Leu-Arg- (SEQ ID NO: 9 in the Sequence Listing). The N-terminal amino acid sequence of the peptide encoded by  
15 the AHAS cDNA of plasmid F2 is Gly-Ser-Leu-Asp-Ala-His-Trp- (SEQ ID NO: 10 in the Sequence Listing). The N-terminal amino acid sequence of the peptide encoded by the AHAS cDNA of the F3 plasmid is Gly-Ser-Val-Glu-Pro-Phe-Phe- (SEQ ID NO: 11 in the Sequence Listing). The N-terminal Gly-Ser- for all three peptides are remnants of the thrombin cleavage.

#### *Expression of the Arabidopsis Large and Small Subunits Proteins of AHAS*

25 15 DH5 $\alpha$ -competent cells (Gibco BRL) were transformed with large subunit plasmids pAC774 and pAC786, as well as small subunit plasmids F1, F2, and F3. Cells were thawed on ice. 1  $\mu$ l of a 1:5 dilution of the plasmid DNA was added to 75  $\mu$ l of cells, which then sat on ice for 30 minutes. Cells were heat shocked in a 42°C water bath for 90 seconds and then put on ice for two minutes. 800  $\mu$ l of Luria-Bertani medium (LB) was added to each tube containing trans-  
30 formed cells which were then grown for 1 hour in a 37°C shaker. Cells were centrifuged for 2 minutes and excess medium was aspirated. The cell pellet was resuspended in 100  $\mu$ l of LB and plated on LB containing 100  $\mu$ g of carbenicillin overnight at 37°C. Single colonies were inoculated into 50 ml of LB medium containing 375  $\mu$ g/ml carbenicillin and grown overnight in a 37 °C shaker. 700  $\mu$ l aliquots were taken and added to 300  $\mu$ l of 50% glycerol, and were then frozen in  
40 25 liquid nitrogen and kept at -80°C for cell stocks.

#### *Purification of the Large Subunit AHAS Gene*

45 An overnight 50 ml culture of transformed *E. coli* harboring the large subunit gene in the pGEX-2T expression vector was inoculated into 1 liter of 2X YT with 2% glucose, 375  $\mu$ g/ml Carbenicillin. Cells were grown for 5 hours in a 37°C shaker/incubator and then induced with 0.1 mM IPTG and then placed in a 30°C shaker for another 2.5 hours. Cells were harvested by centrifugation in a JA10 rotor at 8000 rpm at 4°C for 10 minutes. Cells were stored as a pellet at  
50 -20°C until purification.

5 The cell pellet from 1 liter of cell culture was resuspended in 10 ml of MTPBS (150 mM NaCl, 16 mM Na<sub>2</sub>HPO<sub>4</sub>, 4 mM NaH<sub>2</sub>PO<sub>4</sub>), pH 7.3 (Smith and Johnson *Gene* 67: 31-40, 1998) and 100 µg/ml of lysozyme was added. The suspension was adjusted to 5 mM dithiothreitol. Triton X-100 was added to a final concentration of 1% by addition of a 20% Triton X-100 in  
10 5 MTPBS solution. The cells were shaken gently at 30°C for 15 minutes and were then cooled to 4°C on ice and sonicated for 8-10 seconds using a microtip probe, duty cycle 70%, output control at maximum for the microtip pulser. The sonicate was centrifuged two times in a J20 rotor, 17,000 rpm, 4°C, 10 minutes, to remove insoluble material. Lysate was added to 150 mg (dry weight) of glutathione agarose (equilibrated and hydrated in MTPBS) and inverted for 30 minutes  
15 at 4°C. Agarose was settled by centrifugation at 500 rpm for 5 minutes and was washed with ice cold MTPBS by repeated centrifugation cycles until the A<sub>280</sub> of the wash matched that of  
20 MTPBS. Agarose was transferred to an appropriate column for elution. Fusion proteins were eluted with 50 mM Tris-HCl, 5 mM reduced glutathione into 1 ml fractions. The A<sub>280</sub> of each fraction was checked for protein content and appropriate fractions (A<sub>280</sub> > 0.100) were pooled.  
25 15 To the pooled sample were added 5 units of bovine thrombin per ml of protein solution. The sample was dialyzed against MTPBS, 3 mM dithiothreitol for 15 hours at room temperature to allow proteolytic cleavage of the fusion protein and removal of Tris-HCl and reduced glutathione.  
30 Dialyzed sample was passed twice more through equilibrated glutathione agarose to remove the cleaved GST protein and to remove uncut fusion protein. Purified samples were stored at 4°C  
20 with or without 0.02% sodium azide.

#### *Purification of Small Subunit Protein of AHAS*

35 Transformed DH5 cells were cultured and harvested in a manner similar to that used for collecting cells expressing the large subunit of AHAS. The cell pellet from 1 liter of culture was resuspended in 10 ml of STE (150 mM NaCl, 10mM Tris-HCl pH 8.0, 1 mM EDTA), and then  
40 25 100 µg/ml of lysozyme was added. This was put on ice for 15 minutes. Dithiothreitol was added to 5mM and N-lauroylsarcosine was added to a final concentration of 1.5% from a 10% stock in STE. The cells were vortexed for 10 seconds. The lysate was sonicated on ice for 8-10 seconds using a microtip probe, duty cycle 70%, output control at maximum for the microtip pulser. The sonicate was centrifuged two times in a JA20 rotor at 4°C at 17,000 rpm for 10 minutes, to  
45 remove insoluble material. The supernatant was added to 200mg (dry weight) of glutathione agarose (equilibrated and hydrated in MTPBS) and inverted for 20 minutes at 4°C. The agarose  
30 was settled by centrifugation at 500 rpm for 5 minutes, and the supernatant was aspirated. The  
50

5 agarose was washed with ice cold MTPBS by repeated centrifugation cycles until the  $A_{280}$  of the wash was equal to that of MTPBS. The agarose was transferred to an appropriate column for elution. Fusion proteins were eluted with 50 mM Tris-HCL pH 8.0, 10mM reduced glutathione, 5 mM dithiothreitol, and 2% N-octylglucoside, and 1 ml fractions were collected. The  
10 absorbance at 280 nm of each fraction was checked, and appropriate fractions ( $A_{280} > 0.100$ ) were pooled. The sample was dialyzed for 15 hours against MTPBS, 3mM dithiothreitol, at 4°C to remove reduced glutathione, Tris-HCL, and N-octylglucoside. SDS was added to 0.005% from a 1% stock solution in H<sub>2</sub>O. Five units of bovine thrombin were added per ml of protein solution, and the sample was shaken gently at room temperature for 30 - 45 minutes to allow proteolytic  
15 cleavage of the fusion protein. The sample was then immediately passed through an EtractiGel-D detergent affinity column (Pierce) to remove SDS. The cut sample was stored at 4°C or passed twice through re-equilibrated glutathione agarose to remove GST and uncut fusion protein. The purified sample was stored at 4°C.

#### *Determination of protein concentration*

25 15 An aliquot of protein solution was adjusted to 5% trichloroacetic acid and put on ice for 20 minutes to allow precipitation of protein. The aliquot was then centrifuged for 10 minutes at high speed and 4°C to pellet the precipitate. The pellet was resuspended in an equivalent volume of 3% (w/v) sodium carbonate, 0.1N NaOH. A Pierce BCA protein assay was used to determine  
30 the concentration of the protein solution. Bovine serum albumin standards were prepared in 3% sodium carbonate, 0.1N NaOH.

35 AHAS assays. AHAS assays were performed with slight modification as described by Singh *et al.* (Singh *et al.*, 1988). AHAS activity was assayed in 1X AHAS assay buffer (50 mM HEPES pH 7.0, 100 mM pyruvate, 10mM MgCl<sub>2</sub>, 1mM TPP, and 50  $\mu$ M FAD). A final concentration of 1X assay buffer was obtained either by dilution of enzyme extracted in 2X assay buffer  
40 25 or addition of enzyme to make a final concentration 1X AHAS assay buffer. All assays containing imazethapyr and associated controls contained a final concentration of 5% DMSO due to addition of imazethapyr to assay mixtures as a 50% DMSO solution. Assays were performed in a final  
45 volume of 250 $\mu$ L at 37°C in microtiter plates.

### EXAMPLE 3

30 Effects of Dithiothreitol. The effect of dithiothreitol (DTT) in Phosphate Buffered Saline (MTPBS) on AHAS large subunit enzyme activity was measured. Experiments were carried out  
50

5 with or without 3 mM DTT in the purified protein solution of large subunit. The assays were conducted as above. The results are presented in FIGs. 5 and 6. As can be seen in FIG. 5 when compared to FIG. 6, the catalytic activity of the AHAS large subunit is enhanced dramatically by DTT. Moreover, in the absence of DTT, large subunit activity degraded over the 4 day period  
10 (FIG. 6). The *Arabidopsis* large subunit was found to be very stable in the presence of 3 mM DTT over a period of 1 month when stored at 4°C.

Other sulfhydryl reductants were tested, but DTT appeared to both stabilize and activate  
15 the enzyme better than reduced glutathione and  $\beta$ -mercaptoethanol. Due to the activation of the enzyme by DTT all experiments addressing activation of the AHAS large subunit enzyme by the small subunit enzyme were performed either in the absence of DTT or other sulfhydryl reducing  
20 agents or at a constant concentration of 3 mM DTT.

Effects of Bovine Serum Albumin. To determine whether the enhancement of the large  
subunit was specific to the small subunit, another nonspecific control which tested the effects of bovine serum albumin was run. To a fixed amount of large subunit an increasing amount of a  
25 0.25 mg/ml BSA solution was added. The results are shown in FIG. 7. As seen in FIG. 7, the addition of BSA to the test sample caused a slight, but not significant, and a plateaued increase in the catalytic activity of the large subunit protein of AHAS.

#### EXAMPLE 4

30 The following experiments used the small subunit and large subunit proteins purified as described in Example 2 above. The plasmid F1 containing the AHAS small subunit gene from  
20 *Arabidopsis* was expressed in *E. coli* and partially purified. The protein concentration of the sample was determined and aliquots of increasing concentration were added to a constant amount of purified *Arabidopsis* large subunit.

FIG. 8 shows the activation of the wild type *Arabidopsis* AHAS large subunit by addition  
40 25 of the *Arabidopsis* small subunit protein. AHAS assays were carried out as described above and the results shown in FIG. 8 indicate that small subunit protein enhances the level of enzymatic activity of the catalytic large subunit. The activation of the large subunit protein of AHAS is shown for both the wild type large subunit and a herbicide-resistant mutant of the large subunit.

45 In another experiment, a herbicide-resistant mutant of the large subunit enzyme (substitution mutation at position 124, methionine substituted by histidine) was used. The results shown in  
30 FIG. 9 demonstrate that the enzymatic activity of a herbicide-resistant form of the large subunit is also enhanced.  
50



## EXAMPLE 5

The plant transformation vector, pHUWE67 illustrated in FIG. 10 containing the *Arabidopsis* AHAS small subunit genomic DNA was constructed as follows. The pUC19 vector shown in FIG. 1 containing the 5.6 kb genomic fragment of the *Arabidopsis* AHAS small subunit gene (pMSg6) was cut with Sal I. The 5.6 kb fragment containing the entire *Arabidopsis* AHAS small subunit gene (see FIG. 2), including the promoter and introns was separated from the vector by agarose gel electrophoresis. The fragment was cut out of the agarose gel and purified using the QIAquick Gel Extraction Kit (Cat. No. 28706) following the procedures provided by the manufacturer. The *Agrobacterium* based transformation vector, pBIN19, was cut with Sal I. The vector was purified by phenol:chloroform extractions. The purified vector was dephosphorylated by treatment with calf intestinal alkaline phosphatase and re-extracted with phenol:chloroform. The vector and genomic insert were ligated and the ligation mix containing the construct was used to transform *E. coli* strain DH5 $\alpha$ . *E. coli* was selected on kanamycin plates and plasmids were extracted from transformed *E. coli*. The vector construct was verified by generation of a PCR product and sequencing of the product using the sequencing procedures described in Example 1. The vector designated pHUWE67 (FIG. 10), thus contains a 5.6 kb fragment comprising the AHAS genomic DNA containing the AHAS promoter, an Open Reading Frame (ORF) and 3'-terminator fused with the pBIN19 plasmid. This vector construct is used for *Agrobacterium* based transformation of plants using standard techniques.

The plant transformation vectors illustrated in FIGs. 11A-11E are similarly constructed as vector pHUWE67 above, following standard cloning procedures. In FIG. 11A-11E, the vector may comprise an AHAS small subunit cDNA, fragment, genomic fragment or mutant. In FIG. 11B the vector further comprises an AHAS small subunit promoter operably-linked upstream of the gene insert. In this embodiment, the *Arabidopsis* small subunit gene includes the terminator.

In FIG. 11C the vector construct contains the AHAS small and large subunit genes in tandem, with the large subunit gene downstream from the small subunit gene. Both genes are regulated by the AHAS small subunit promoter which is located upstream from the gene inserts. The AHAS large subunit gene is a herbicide resistant mutant allele.

In FIG. 11D the plant expression vector contains the large and small subunit genes under the control of their own promoters. Transcription termination signal is provided by the AHAS small subunit terminator. The AHAS large subunit gene confers resistant to herbicides such as imidazolinone.

5           The vector in FIG. 11E is similar to the vector represented in FIG. 11C. In this vector, however, the AHAS large and small subunit genes are reversed in position in the construct. The AHAS large subunit gene in this example is upstream of the small subunit gene. The AHAS large subunit gene is a herbicide resistant mutant allele.

10           5           Based on the techniques of present invention and following astringent DNA hybridization techniques, homologous AHAS small subunit gene sequences can be obtained from a variety of plant species, such as rice, maize, wheat, barley, and the like. Therefore, these AHAS small subunit gene sequences are also useful in the present vectors and methods for transforming plants.

## Claims

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We claim:

1. An isolated DNA sequence encoding a eukaryotic acetohydroxy-acid synthetase, AHAS small subunit protein.
2. The DNA sequence of claim 1 shown as SEQ ID NO: 1 in the Sequence Listing.
3. The DNA sequence of claim 1 shown as SEQ ID NO: 3 in the Sequence Listing.
4. A DNA sequence of claim 1 which encodes a plant AHAS small subunit protein.
5. A DNA sequence of claim 4 which encodes a dicotyledonous AHAS small subunit protein.
6. The DNA sequence of claim 5 which encodes the AHAS small subunit protein from *Arabidopsis*.
7. A DNA sequence of claim 4 which encodes a monocotyledonous AHAS small subunit protein.
8. A DNA sequence of claim 7 which encodes the AHAS small subunit protein from rice or maize.
9. A plant expression vector comprising a promoter expressible in a plant cell and a DNA sequence encoding a eukaryotic AHAS small subunit protein.
10. A expression vector of claim 9, wherein the promoter is a high level expression plant promoter.
11. A plant expression vector of claim 9, wherein the promoter is the *Arabidopsis* AHAS small subunit promoter.
12. The plant expression vector of claim 11, wherein the DNA sequence encodes the *Arabidopsis* small subunit protein.
13. A plant expression vector of claim 9, further comprising a second promoter expressible in a plant cell and a DNA sequence encoding a eukaryotic AHAS large subunit protein.
14. A plant expression vector of claim 13, wherein the large subunit promoter and DNA sequence encoding a eukaryotic AHAS large subunit protein are derived from a dicotyledonous plant.
15. A plant expression vector of claim 14, wherein the dicotyledonous plant is *Arabidopsis*.
16. A plant expression vector of claim 13, wherein the DNA sequence encodes a variant AHAS large subunit protein which is resistant to herbicides.

5 17. A plant expression vector of claim 16, wherein the *Arabidopsis* DNA sequence encodes a mutant *Arabidopsis* AHAS large subunit protein which is resistant to imidazolinone herbicides.

10 18. A plant expression vector of claim 13, wherein the promoters and the DNA sequences are derived from a dicotyledonous plant.

19. A plant expression vector of claim 13, wherein the promoters and the DNA sequences are derived from a monocotyledonous plant.

15 20. A plant expression vector of claim 19, wherein the DNA sequences are derived from rice or maize.

10 21. A plant expression vector for expressing an AHAS gene in a plant, comprising a promoter and DNA sequence encoding a fusion protein comprising a large subunit and a small subunit of an AHAS protein.

22. A plant expression vector of claim 21, wherein the DNA sequence encoding the small subunit of the AHAS protein is derived from eukaryotes.

25 15 23. The plant expression vector of claim 22, wherein the promoter and the DNA sequences are derived from *Arabidopsis*.

30 24. A plant expression vector comprising in series a promoter expressible in a plant cell; a DNA sequence encoding a transit AHAS large subunit polypeptide; a DNA sequence encoding mature AHAS large subunit protein; a DNA sequence encoding a linker polypeptide; a DNA sequence encoding a eukaryotic AHAS small subunit protein; and a plant terminator sequence.

35 25. A plant expression vector comprising in series a promoter expressible in a plant cell; a DNA sequence encoding a transit AHAS small subunit polypeptide; a DNA sequence encoding an AHAS small subunit protein; a DNA sequence encoding a linker polypeptide; a DNA sequence encoding a AHAS large subunit polypeptide and a plant terminator.

40 26. A plant expression vector of claim 24 or 25, wherein the promoter, the DNA sequence encoding the transit AHAS large subunit protein and the DNA sequence encoding the mature AHAS large subunit protein are derived from a dicotyledonous plant.

45 27. The plant expression vector of claim 24 or 25, wherein the promoter, the DNA sequence encoding the transient AHAS large subunit protein and the DNA sequence encoding the mature AHAS large subunit protein are derived from *Arabidopsis*.

5 28. A plant expression vector of claim 24 or 25, wherein the promoter, the DNA sequence encoding the transit AHAS large subunit protein and the DNA sequence encoding the mature AHAS large subunit protein are derived from a monocotyledonous plant.

10 29. The plant expression vector of claim 24 or 25, wherein the promoter, the DNA sequence encoding the transit AHAS large subunit protein and the DNA sequence encoding the mature AHAS large subunit protein are derived from rice or maize.

15 30. An isolated eukaryotic AHAS small subunit protein.

31. The protein of claim 30 which has the amino acid sequence corresponding to SEQ ID NO: 2 in the Sequence Listing.

10 32. A composition comprising a purified eukaryotic AHAS small subunit protein.

20 33. The composition of claim 32, wherein the AHAS small subunit protein is an *Arabidopsis* AHAS small subunit protein.

25 34. A method for creating a transgenic plant which is resistant to herbicides, comprising transforming a plant with a plant expression vector comprising a DNA sequence encoding a eukaryotic AHAS small subunit protein.

30 35. The method of claim 34, wherein the expression vector further comprises a high level expression plant promoter and a DNA sequence encoding a eukaryotic AHAS large subunit protein.

20 36. The method of claim 35, wherein the promoter is a eukaryotic AHAS small subunit promoter and the large subunit DNA sequence encodes a eukaryotic herbicide resistant mutant of an AHAS large subunit protein.

35 37. The method of claim 36, wherein the promoter and DNA sequence encoding the AHAS large subunit protein are derived from *Arabidopsis*.

40 38. A method for imparting herbicide resistance to a plant cell, comprising co-transforming the plant cell with a first plant expression vector comprising a first plant expressible promoter, and a DNA sequence encoding the large subunit of the AHAS protein and a second plant expression vector comprising a second plant expressible promoter and a DNA sequence encoding the small subunit of an eukaryotic AHAS protein.

45 39. A plant cell produced by the method of claim 38.

30 40. A method for enhancing herbicide resistance of a transgenic plant which expresses a gene encoding an AHAS large subunit protein comprising transforming the transgenic plant with a DNA sequence encoding a eukaryotic small subunit AHAS protein.

5 41. A transgenic plant produced by the method of claim 40.

42. A transgenic plant produced by the method of claim 40, wherein the plant exhibits an elevated resistance to imidazolinone herbicides as compared to other AHAS inhibiting herbicides.

10 43. The transgenic plant of claim 42, wherein the other AHAS inhibiting herbicides are  
5 selected from the group consisting of sulfonylureas, triazolopyrimidines, pyrimidyl-oxy-benzoic acids, sulfamoylureas and sulfonylcarboxamides.

15 44. A method for enhancing the herbicide resistance in the progeny plants of a plant which comprises crossing the plant with a transgenic plant whose genetic complement comprises a sequence encoding a herbicide-resistant mutant of the large subunit of a eukaryotic AHAS  
10 protein, and a DNA sequence encoding the small subunit of a eukaryotic AHAS protein; and selecting for those progeny plants which exhibit herbicide resistance.

20 45. The method of claim 44, wherein crossing the plant with a transgenic plant is accomplished by sexual reproduction.

46. A progeny plant produced by the method of claim 44.

25 15 47. A progeny plant of claim 44 whose genetic complement contains a DNA sequence encoding the eukaryotic small subunit AHAS protein.

30 48. A herbicide resistant derivative plant of a plant of claim 44 whose genetic complement comprises the DNA sequence encoding the herbicide resistant mutant of the large subunit of the eukaryotic AHAS protein and the DNA sequence encoding the small subunit of the eukaryotic  
20 AHAS protein.

35 49. A plant which is a derivative of the plant of claim 48 and which exhibits a tolerance to the imidazolinone herbicides.

40 50. A transgenic plant whose genetic complement comprises a plant expressible gene comprising a promoter for expression in plants, a DNA sequence encoding a fusion protein  
25 comprising a large subunit and a small subunit of a eukaryotic AHAS protein and a terminator sequence which functions in plant cells.

51. A plant derivative of the plant of claim 49 whose genetic complement contains a DNA sequence encoding a eukaryotic small subunit AHAS protein.

45 52. A transgenic plant of claim 46 or 47 which is a dicotyledonous plant.

30 53. A transgenic plant of claim 46 or 47 which is a monocotyledonous plant.

5 54. A method for identifying mutations in the plant AHAS gene which confer resistance to herbicides, comprising exposing an organism to a herbicide compound, which organism possesses a heterologous vector comprising an AHAS small subunit protein gene.

10 55. The method of claim 54, wherein the vector further comprises an AHAS large subunit protein gene.

56. The method of claim 54, wherein the AHAS small subunit gene has a DNA sequence derived from *Arabidopsis*.

15 57. A screening method for identifying herbicides which effect AHAS enzymatic activity, comprising exposing an organism to a compound, said organism possessing a heterologous expression vector comprising an AHAS small subunit protein gene, and determining the effects of the herbicide

20 58. A screening method for identifying a mutation which alters allosteric feedback inhibition of the AHAS enzyme, comprising:

25 15 transforming a microbial strain which is deficient in AHAS enzyme activity with a plasmid expression vector comprising a mutant plant AHAS small subunit gene;

screening the microbial strain in minimal medium in the presence of one or two branched chain amino acids, and

30 identifying the microbial strains which grow in said medium.

59. An isolated DNA sequence comprising nucleotide sequences 1-757 of SEQ ID NO:3.

20 60. A heterologous plant expression vector comprising the isolated DNA sequence of claim 59.

35 61. The plant expression vector according to claim 60, comprising a heterologous DNA sequence.

40 25 62. A method for expressing a heterologous DNA sequence in a plant cell comprising transforming the plant cell with the expression vector of claim 61.

63. A recombinant plant cell comprising the DNA sequence of claim 59.

64. The recombinant plant cell of claim 62.



*Arabidopsis* AHAS Small Subunit Gene

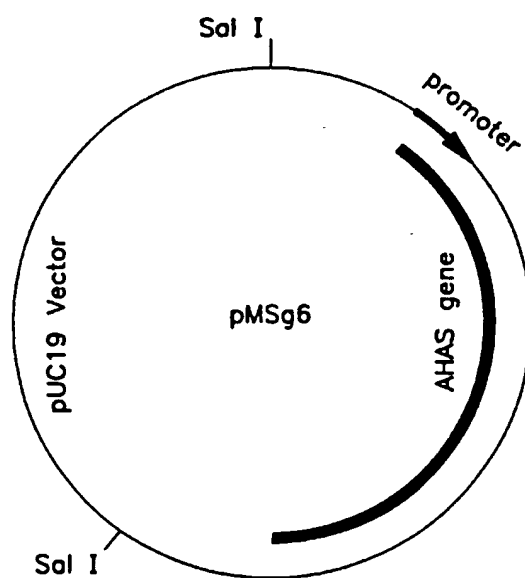


FIG. 1

2 / 9

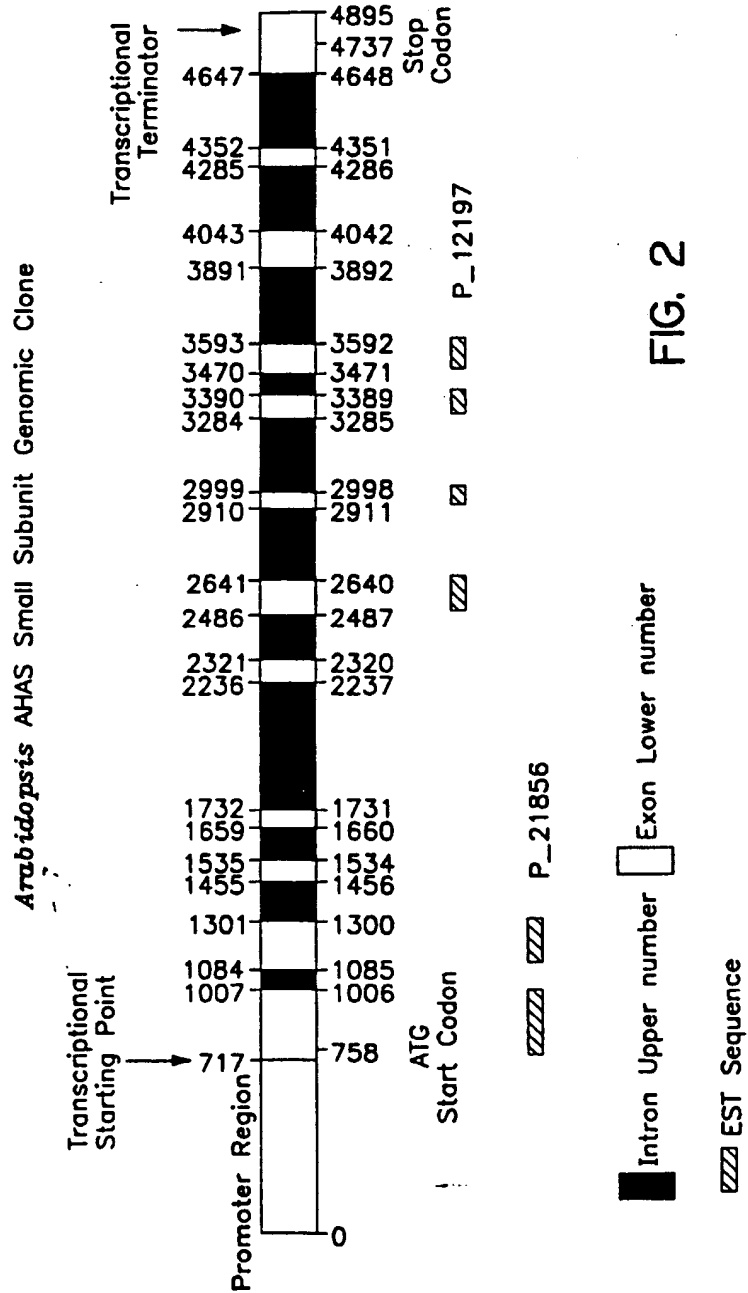


FIG. 2

*Arabidopsis* AHAS Small Subunit Expression Vectors  
pHUWE82 and pHUWE83

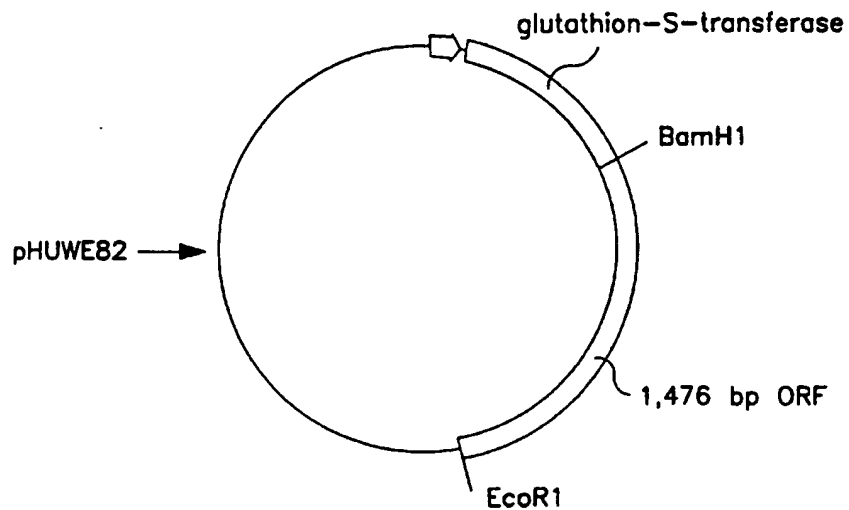


FIG. 3

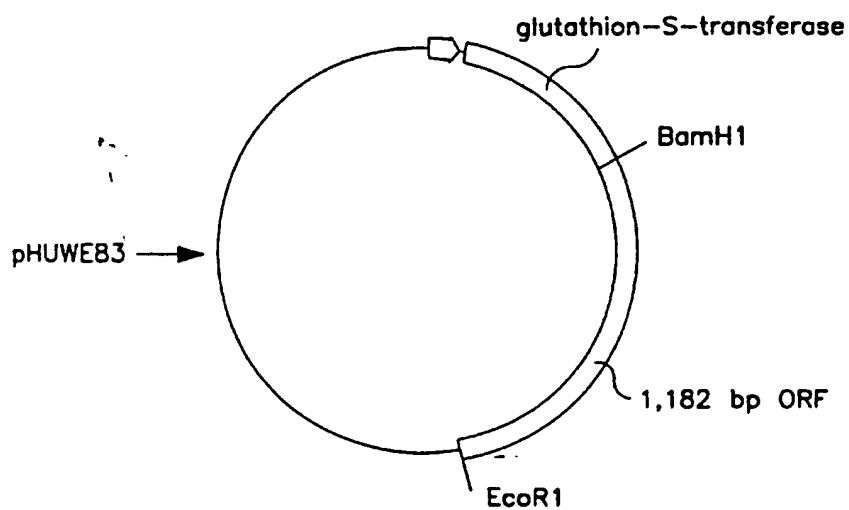


FIG. 4

4 / 9

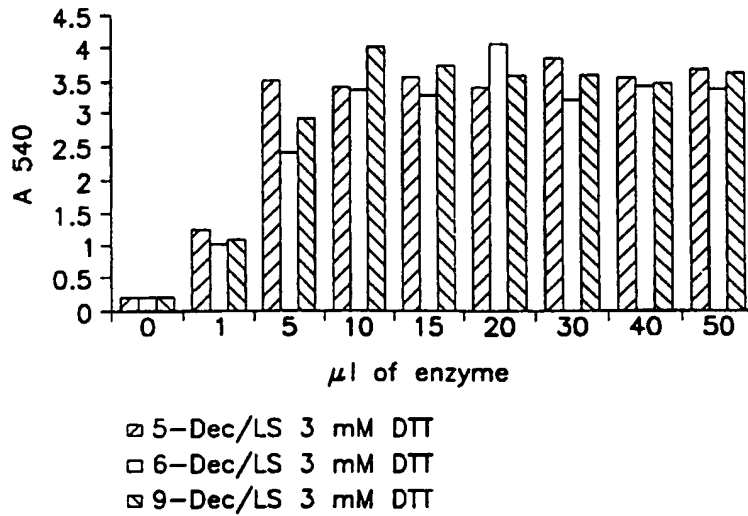
Activity of Ar-AHAS Large Subunit in MTPBS;  
3mM DTT over time

FIG. 5

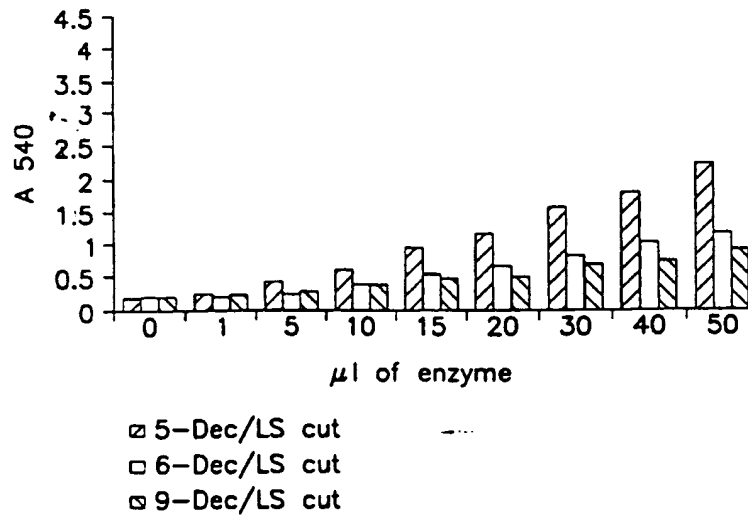
Ar-AHAS Large Subunit in MTPBS  
over time

FIG. 6

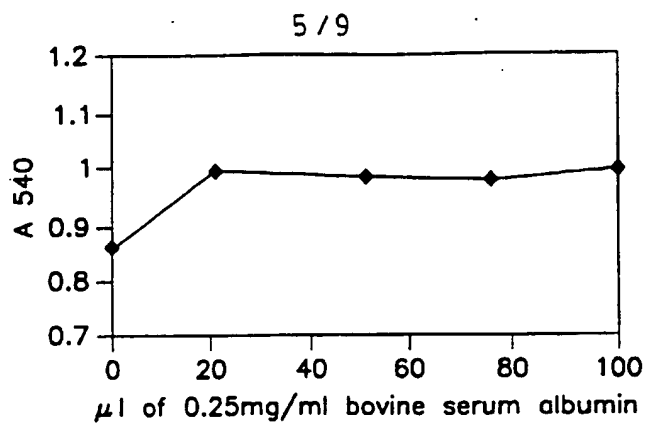


FIG. 7

Activation of the Wild Type Large Subunit of  
*Arabidopsis* AHAS by the *Arabidopsis* AHAS Small Subunit

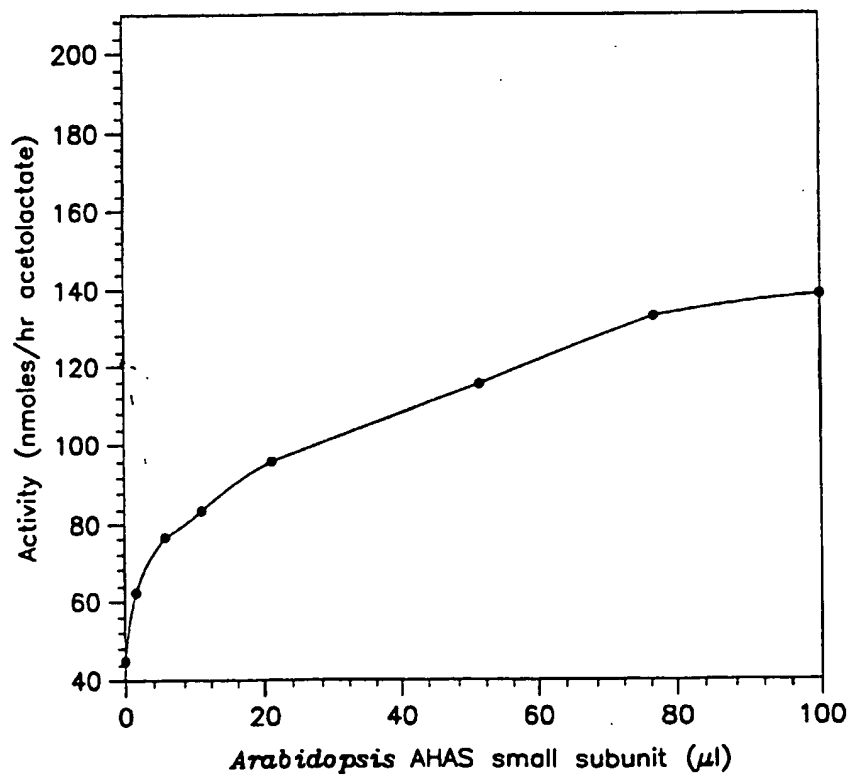


FIG. 8

Activation of the Herbicide Resistant Mutant (Met124His) Large Subunit of *Arabidopsis* AHAS by the *Arabidopsis* AHAS Small Subunit

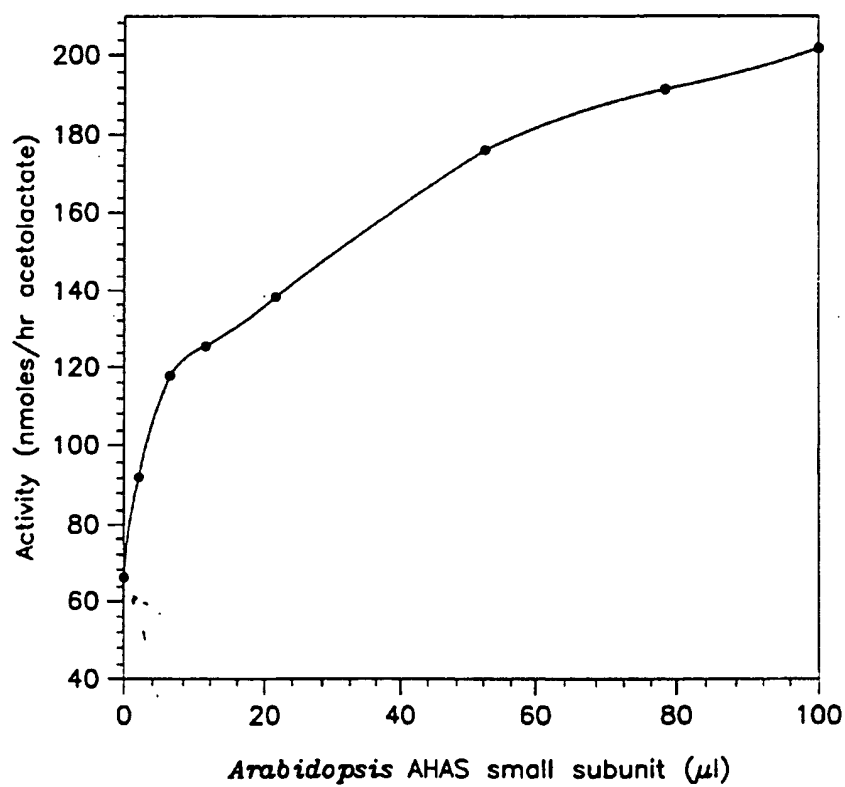


FIG. 9

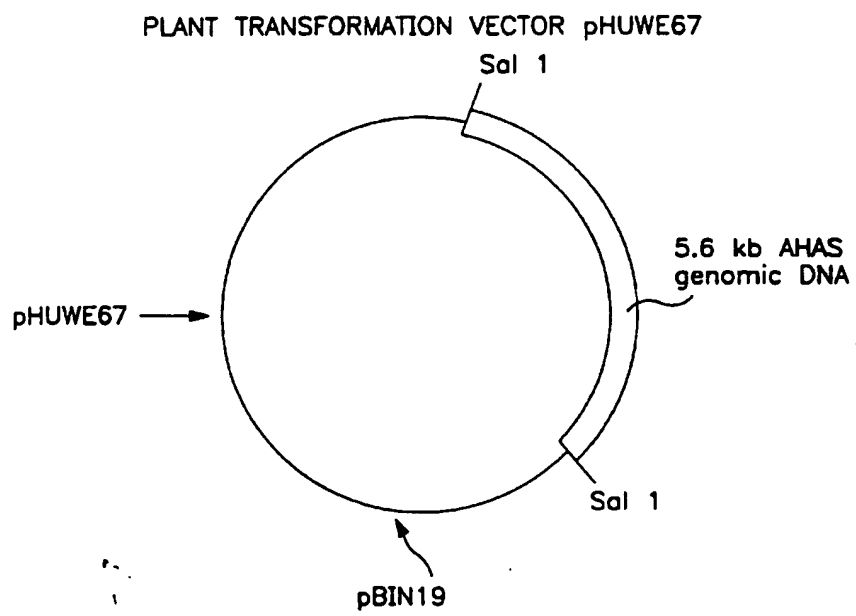


FIG. 10

# PLANT EXPRESSION VECTORS

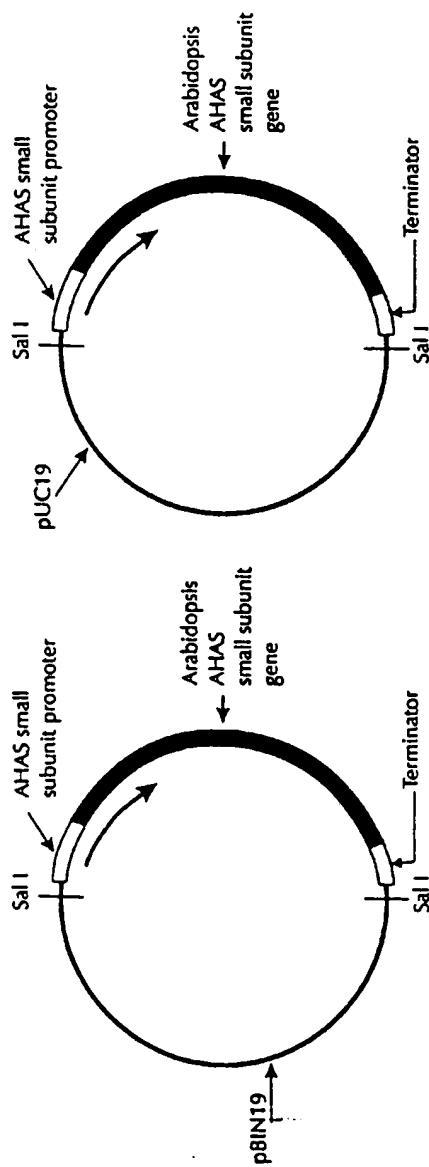


FIG. 1B

FIG. 1A



PLANT EXPRESSION VECTORS

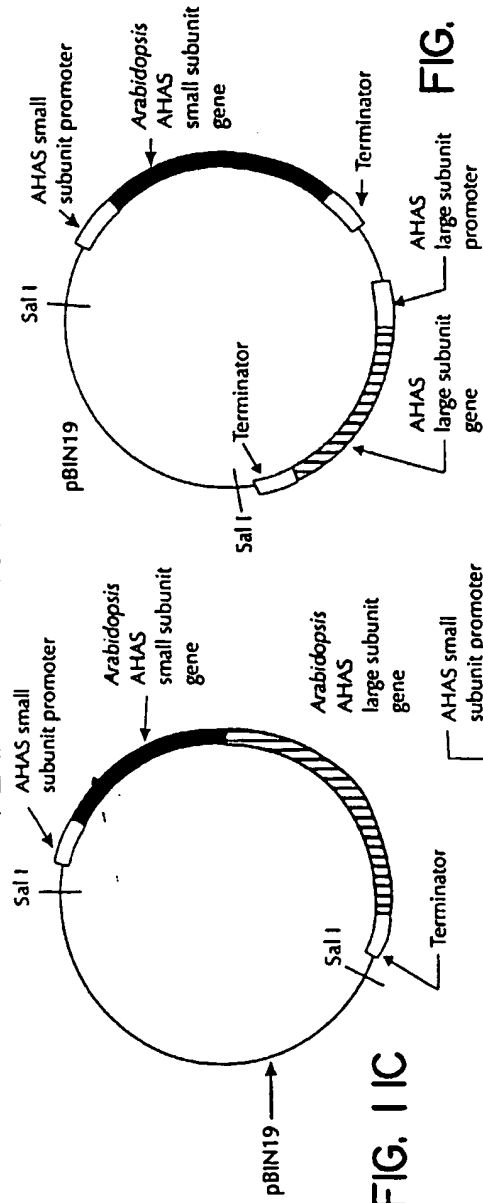


FIG. 1 IC

FIG. 1 ID

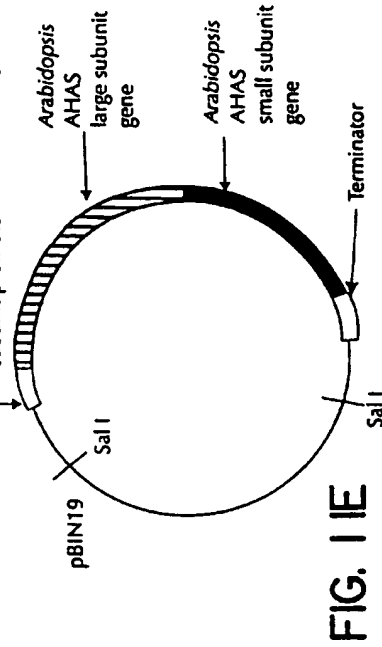


FIG. 1 IE

## SEQUENCE LISTING

<110> Kakefuda, Genichi  
Costello, Colleen  
Sun, Ming  
Hu, Weiming

<120> Genes and Vectors for Conferring Herbicide Resistance  
in Plants

<130> 1102319-0003

<140>

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## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification:</b> C12N 15/82, A01H 5/00, C12N 5/10, C12N 9/88, C12N 15/60	<b>A2</b>	<b>(11) International Publication Number:</b> WO 00/26390 <b>(43) International Publication Date:</b> 11 May 2000 (11.05.2000)
<b>(21) International Application Number:</b> PCT/US99/25452 <b>(22) International Filing Date:</b> 28 October 1999 (28.10.1999) <b>(30) Priority Data:</b> 60/106,239 29 October 1998 (29.10.1998) US <b>(60) Parent Application or Grant</b> AMERICAN CYANAMID COMPANY [/]; (). KAKEFUDA, Genichi [/]; (). COSTELLO, Colleen [/]; (). SUN, Ming [/]; (). HU, Weiming [/]; (). SPRULL, W., Murray, ; ().		<b>Published</b>
<b>(54) Title: GENES AND VECTORS FOR CONFERRING HERBICIDE RESISTANCE IN PLANTS</b> <b>(54) Titre: GENES ET VECTEURS SERVANT A CONFERER UNE RESISTANCE AUX HERBICIDES AUX PLANTES</b>  <b>(57) Abstract</b> <p>Genomic and cDNA sequences and plant expression vectors encoding for an eukaryotic AHAS small subunit protein are disclosed. The DNA sequences and vectors are used to transform plants to produce transgenic plants which possess elevated levels of tolerance or resistance to herbicides, such as imidazolinones.</p> <b>(57) Abrégé</b> <p>L'invention concerne des séquences génomiques et des séquences d'ADNc, ainsi que des vecteurs d'expression de plantes codant pour une protéine de petite sous-unité eucaryote AHAS. On utilise ces séquences d'ADN et ces vecteurs afin de transformer des plantes pour produire des plantes transgéniques possédant des niveaux élevés de tolérance ou de résistance aux herbicides, tels que les imidazolinones.</p>		